



PARKER & PARSLEY AUSTRALASIA LIMITED

PEP 108 OTWAY BASIN - VICTORIA

# MYLOR #1 WELL COMPLETION REPORT

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Well Completion Report vol. 2 Mylor-1 Will(2)

## APPENDIX 3

PETROLEUM DIVISION

#### **APPENDIX**

3. Palynological Report

#### MORGAN PALAEO ASSOCIATES

PALYNOLOGICAL/PETROLEUM GEOLOGICAL CONSULTANTS

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PALYNOLOGY OF BRIDGE MYLOR-1

ONSHORE OTWAY BASIN, VICTORIA

BY

**ROGER MORGAN** 

for **BRIDGE OIL** 

**Sept 1994** 

OTW.RPMYLOR



#### PALYNOLOGY OF BRIDGE MYLOR-1

#### ONSHORE OTWAY BASIN, VICTORIA

#### BY

#### **ROGER MORGAN**

	CONTENTS	PAGI	<u>.</u>
I	SUMMARY		3
II	INTRODUCTION		4
Ш	PALYNOSTRATIGRAPHY		5
IV	REFERENCES		9
	FIGURE 1 : CRETACEOUS REGIONAL FRAMEWORK	X, OTWAY I	BASIN
	FIGURE 2 MATURITY PROFILE : MYLOR-1		

#### I SUMMARY

- 1388.0m(swc), 1391.0m(swc): upper *apoxyexinus* Zone (upper *cretacea* Dino Zone): late Santonian: very nearshore marine: immature for hydrocarbons: seen in Paaratte and Belfast Formations and equivalents
- 1500.0m(swc), 1515.0m(swc): middle apoxyexinus Zone (lower cretacea Dino Zone): mid Santonian: nearshore marine: immature to early marginal mature for oil, immature for gas/condensate: seen in the Paaratte and Belfast Formations and equivalents
- 1650.0m(swc), 1658.0m(swc): lower *apoxyexinus* Zone: early Santonian: nearshore marine: marginally mature for oil, immature for gas/condensate: seen in the Belfast and Flaxmans Formations and equivalents
- 1672.0m(swc): mawsonii Zone: Turonian-Coniacian: brackish lagoon: marginally mature for oil, immature for gas/condensate, algal rich: seen in the Belfast and Flaxmans Formations and equivalents
- 1758.5m(swc), 1763.0m(swc): extremely lean and zonally indeterminate, apparently non-marine or slightly brackish: marginally mature for oil, immature for gas/condensate
- 1833.0m(swc): apparently *paradoxa* Zone, but the usual markers are extremely rare as in the sandy Eumeralla facies offshore: mid Albian: slightly brackish lagoon: borderline mature for oil, borderline marginally mature for gas/condensate: usually Eumeralla Formation.

#### II INTRODUCTION

After well completion, ten sidewall cores were submitted for detailed study. All results are summarised herein.

Palynomorph occurrence data are shown as Appendix I and form the basis for the assignment of the samples to five spore-pollen and dinoflagellate units of Santonian to Albian age. Specimen counts were made on all assemblages and expressed in the raw data as percentages. The marine fossils are presented as a percentage of total fossils (marine plus non-marine) in the raw data in Appendix 1 as an expression of marineness.

The Cretaceous spore-pollen zonation is essentially that of Dettmann and Playford (1969), but has been significantly modified and improved by various authors since, and most recently discussed in Helby et al (1987), as shown on Figure 1. The Late Cretaceous zonation has been modified by Morgan (1992) in project work and recent offshore drilling for BHPP and partners.

Maturity data was generated in the form of Spore Colour Index, and is plotted on Figure 2 Maturity Profile of Mylor-1. The oil and gas windows on Figure 2 follow the general consensus of geochemical literature. The oil window corresponds to spore colours of light-mid brown (Staplin Spore Colour Index of 2.7) to dark brown (3.6). These correspond to Vitrinite Reflectance values of 0.6% to 1.3%. Geochemists argue variations on kerogen type, basin type and basin history. The maturity interpretation is thus open to reinterpretation using the basic colour observations as raw data. However, the range of interpretation philosophies is not great, and probably would not move the oil window by more than 200 metres.

		MICRO-		LITHOSTRATIGRAPHY
AGE	SPORE POLLEN	PLANKTON		
1984	ZONES	ZONES		. OFFSHORE
MAASTRICHTIAN	T.longue	Ldruggii	<b>}</b>	TIMBOON SANDSTONE
	T.IIIei,	1.korojoneme		~~~
CAMPANIAN	N.senectus Nothor		quosa	SHERBROOK GROUP  (thin sendstones)
SANTONIAN	T	l.cretaceum	BROOK	
CONIACIAN		O.porifora	HERE	BELL'AST MUDSTONE
TURONIAN	C.triplex	C.striatoconus	3	
CENOMANIAN	A.distocarinatus	P.infusorioides D.multispinum		FLAXMANS FORMATION  WAARE SANDSTONE
ALBIAN	P.pannosus Up.C.paradoxa Low.C.paradoxa C.striatus	<del>, , , , , , , , , , , , , , , , , , , </del>	<b>}</b>	middle SHeathfield Member EUMERALLA FORMATION (EGally (acked)
APTIAN	Upper C.hughesi Low .C.hughesi	,		lower  Windermerg M Coaly factor  (KGTANTER D COAL)
BARREMIAN	upper F. wonthaggiensis	<del></del> 1	SROUP	Crayfish C (Laira Fm)
HAUTERIVIAN	lower F. wonthaggiensis		YAWTO	H SUBGRO
VALAHGINIAN				Pretty H
BEŖRIASIAN	C. australiersis			AAAA
		ייסטיינייניסטיי	\ \tilde{5}	NO A COMPANY

FIGURE 1. CRETACECUS REGIONAL FRAMEWORK, OTWAY BASIN

		DE	•	im	matu	r e		i	ture		dry	gas /		GAS/ CONDENSATE
A	ZC	РТН		immat	ure		marg -inal	mature	, t	os	t m	ature		OİL
AGE	ZONE	(tho				yellow	light	brow mid	n \ dar	$\frac{1}{k}$		blac	ck	COLOUR
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ı u-Co	maws	-	٠		,		•							
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		_												
		2.5						1						

FIGURE 2 MATURITY PROFILE : MYLOR-I

#### III PALYNOSTRATIGRAPHY

#### A 1388.0m(swc), 1391.0m(swc): upper apoxyexinus Zone (upper cretacea dino Zone)

Assignment to the upper *Tricolporites apoxyexinus* Zone of late Santonian age is indicated at the top by the absence of younger markers (such as *Nothofagidites senectus*) and at the base by consistent but rare *Amosopollis cruciformis* (1% or less). *Falcisporites* and *Microcachryidites* are common with *Australopollis obscurus*, *Cyathidites*, *Osmundacidites*, *Podosporites*, *Proteacidites* spp and *Vitreisporites* frequent. Angiosperms are rare with *Tricolpites gillii* and *Tricolporites apoxyexinus* seen.

Assignment to the upper *Isabelidinium cretacea* dinoflagellate Zone is indicated at the top by youngest *Amphidiadema denticulata* and *Isabelidinium belfastense rotundata* and at the base by oldest *Isabelidinium belfastense belfastense*. Dinoflagellates are all rare but *Heterosphaeridium heteracanthum* is the most frequent. *Trithyrodinium* spp and *Odonotochitina* spp are striking elements.

Environments are very nearshore marine, as shown by the low dinoflagellate content (3% and 11% downhole) and moderate diversity.

These features are normally seen in the Paaratte Formation (including the Skull Creek and Nullaware Members) and Belfast Mudstone and their equivalents.

They occur in the Sherbrook Group above the Shipwreck Group offshore.

Yellow to light brown spore colours indicate immaturity for hydrocarbon generation.

#### B 1500.0m(swc), 1515.0m(swc): middle apoxyexinus Zone (lower cretacea dino Zone)

Assignment to the middle *T. apoxyexinus* Zone is indicated at the top by the downhole influx of *A. cruciformis* and at the base by the absence of older indicators. *Cyathidites* and *Falcisporites* were common, with *Dilwynites*, *Podosporites*, *Proteacidites* and *Vitreisporites* frequent. *T. gillii* occurs only at 1500m. Very rare elements include *Aequitriradites* spp, *Australopollis obscurus*, *Foraminisporis wonthaggiensis*, and *Phyllocladidites mawsonii*.

Assignment to the lower *I. cretacea* dinoflagellate Zone is indicated at the top by the absence of younger markers and at the base by oldest *I. cretacea*.

Heterosphaeridium spp are frequent to common with rare taxa including Odontochitina spp, O. porifera and Trithyrodinium spp.

Nearshore marine environments are indicated by the low dinoflagellate content (19 and 23% downhole), and their moderate diversity.

These features are normally seen in th Paaratte Formation and Belfast Mudstone and their equivalents. They occur in the Sherbrook Group above the Shipwreck Group offshore.

Light brown to yellow spore colours indicate early marginal maturity for oil and immaturity for gas/condensate.

#### C 1650.0m(swc), 1658.0m(swc): lower apoxyexinus Zone

Assignment to the lower *T. apoxyexinus* Zone is indicated at the top by the major downhole influx of *A. cruciformis* (9-14%) and at the base by the base of the *A. cruciformis* acme and absence of older markers. *Dilwynites granulatus* is very common, with *Falcisporites* and *A. cruciformis* common. Rare elements include *A. obscurus*, *Clavifera triplex* and *P. mawsonii*.

Dinoflagellates are rare and lack the published zone markers. However, youngest Aptea sp (1658m) Chlamydophorella ambigua and consistent Circulodinium deflandrei (1650m) usually occur within the lower apoxyexinus Zone and so are consistent. Heterosphaeridium spp C. deflandrei and Spiniferites spp are the most frequent taxa. A small undescribed acritarch informally called Rectanguladinium sp occurs only in these two samples and may have future biostratigraphic significance.

Nearshore marine environments are indicated by the low to moderate dinoflagellate content (17 and 34% downhole) and their moderate diversity.

These features are normally seen in the Belfast Mudstone and Flaxmans Formations and their equivalents. They occur in the upper Shipwreck Group offshore.

Light brown spore colours indicate marginal maturity for oil and immaturity for gas/condensate.

#### D 1672.0m(swc): mawsonii Zone

Assignment to the *Phyllocladidites mawsonii* Zone of Coniacian-Turonian age is indicated at the top by youngest *Appendicisporites distocarinatus* and at the base by oldest *P. mawsonii*. *Cyathidites* spp are abundant with *Microcachryidites* and *Vitreisporites* common, and *Dilwynites* and *Falcisporites* frequent.

Dinoflagellates are very scarce, with only a few single specimens seen. Brackish environments are therefore indicated with the dinoflagellates comprising less than 1% of the assemblage and of very low diversity. Freshwater algae (*Botryococcus*) are abundant (17%) and suggest lacustrine environments. The rare dinoflagellates however argue for minor saline influence and a tidal lagoon or coastal lake are likely.

These features are normally seen in the Belfast and Flaxmans Formations and their equivalents. They occur in the lower part of the upper Shipwreck Group offshore, including the reservoir section in the Minerva field.

Light brown spore colours indicate marginal maturity for oil and immaturity for gas/condensate.

#### E 1758.5m(swc), 1763.0m(swc): lean and indeterminate

These two samples are extremely lean and dominated by inertinite and cuticle fragments. The relatively rare age diagnostic taxa were not seen and they cannot be assigned to any zone. Falcisporites are abundant with Cyathidites and Odmundacidites common and Retitriletes austroclavatidites frequent. Permian reworking is prominent.

At 1758.5m, saline markers are absent and freshwater algae (*Botryococcus*) comprises 4% of the assemblage. This suggests freshwater environments with lacustrine influence. However, too little material is available to deny the possibility of marine influence.

At 1763m, spiny acritarchs (3%) indicate slight saline influence. Freshwater algae (*Botryococcus* and *Schizosporis*) are present. Slightly brackish environments are therefore indicated.

Light brown spore colours indicate marginal maturity for oil and immaturity for gas/condensate.

#### F 1833.0m: apparently paradoxa Zone

Assignment to the *Coptospora paradoxa* Zone of mid Albian age is suggested at the top by the absence of younger markers (such as *Phimopollenits pannosus*) with youngest *C. paradoxa* and at the base by oldest *C. paradoxa*. However, the assemblage lacks rich heavy spore assemblages normally seen in Eumeralla Formation claystones, as apparently occurs in recent offshore drilling nearby. Instead, the assemblage is very bland, dominated by saccates and smooth spores, and lacking the usual diverse ornamented spore assemblage. As a result, this assemblage might be as young as the Cenomanian *distocarinatus* Zone.

In five microscope slides examined, I saw only one specimen of *C. paradoxa*, two specimens of *Foraminisporis asymmetricus* and no *Crybelosporites striatus*. These are usually all consistent in the Eumeralla Formation.

Brackish influence is slight but seen in very rare spiny acritarchs amongst the dominant and diverse spores and pollen. Freshwater algae (*Botryococcus*) are frequent (5%) suggesting lakes. Brackish nearshore lagoons seem likely.

The *paradoxa* Zone is usually seen in the Eumeralla Formation. However, the tentative nature of the assignment means that palynology cannot definitively confirm penetration of the Eumeralla. This must rest on lithological or other criteria. Palynologically, it is not impossible that the Eumeralla was not drilled.

#### IV REFERENCES

- Dettmann ME and Playford G (1969) Palynology of the Australian Cretaceous: a review In Stratigraphy and Palaeontology. Essays in honour of Dorothy Hill, KSW Campbell ED. ANU Press, Canberra 174-210
- Helby RJ, Morgan RP and Partridge AD (1987) A palynological zonation of the Australian Mesozoic In Studies in Australian Mesozoic Palynology Assoc. Australas. Palaeontols. Mem 4 1-94
- Morgan RP (1992) Overview of new cuttings based Late Cretaceous correlations, Otway Basin, Australia unpubl. rept. for BHPP.

#### MYLOR #1

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C L I E N T: BRIDGE OIL LTD

WELL: MYLOR #1

F I E L D / A R E A: ONSHORE OTWAY BASIN,

PEP 108, VICTORIA

A N A L Y S T: ROGER MORGAN

D A T E: SEPTEMBER '94

N O T E S: ALL DEPTHS IN METRES. ALL FIGURES ARE PERCENTAGES.

X MEANS THAT SPECIES IS VERY RARE AND OCCURRED OUTSIDE GRAIN COUNT. IN UNCOUNTED SAMPLES A=ABUNDANT, C=COMMON, F=FREQUENT.

RANGE CHART OF OCCURRENCES BY % & LOWEST APPEARANCE: grouped

1388.0 SWC 3 1391.0 SWC 1500.0 SWC 1515.0 SWC 1650.0 SWC 1658.0 SWC 1672.0 SWC 1758.0 SWC 1758.0 SWC 1763.0 SWC	
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	11 CRIBROPERIDINIUM SPP
, , , <sub> </sub>	13 FLORENTINIA DEANEI
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1 . 27 %	15 HETEROSPHAERIDIUM SOLIDA
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, , , <sub> </sub> , , , , , ,	17 KIOKANSIUM RECURVATUM
· · · · • • • × · · ·	18 NUMMUS SP
× . ×	19 ODONTOCHITINA OPERCULATA
××	
	23   SPINIFERITES FURCATUS/RAMOSUS
, , , , <u>,</u> , , , , ,	24 TRICHODINIUM
××	25 CALLACISPHAERIDIUM ASYMMETRICUM
×	26   CRIBROPERIDINIUM EDWARDSII 
×	27 CYCLONEPHELIUM MEMBRANIPHORUM
	28    HYSTRICHODINIUM PULCHRUM 
	29      29   ISABELIDINIUM SP 
	30    CASSICULOSPHAERIDIA GIANT 
×	31   FROMEA FRAGILIS 
, , , , , , <u>,</u> , , ,	32   GILLINIA HYMENOPHORA    "
× × × ×	33   ISABELIDINIUM CRETACEUM 
	34  ODONTOCHITINA COSTATA
	35   ODONTOCHITINA CRIBROPODA 
	36   ODONTOCHITINIA PORIFERA 
×	37   PALAEOPERIDINIUM CRETACEUM
	TRITHYRODINIUM PUNCTATE    38   TRITHYRODINIUM PUNCTATE

1388.0 SWC 3 1391.0 SWC 1500.0 SWC 1515.0 SWC 1650.0 SWC 1658.0 SWC 1672.0 SWC 1758.0 SWC 1763.0 SWC 1763.0 SWC	# H H H H H H H H H H H H H H H H H H H	
×××× · · · · · · · · · · · · · · · · ·	39	TRITHYRODINIUM THICK RETICULATE
	40	CHATANGIELLA VICTORIENSIS
	41	EUCLADINIUM MADURENSE
	42	HETEROSPHAERIDIUM CF LATEROBRACHIUS
	43 H	ODONTOCHITINIA TRIANGULARIS
	44	PARALECANIELLA
	45      45	CANNINGIA SPP
	46      46	HETEROSPHAERIDIUM ROBUSTA
	47      47	ISABELIDINIUM BELFASTENSE
	48    	ODONTOCHITINA STUBBY
	49      	TRITHYRODINIUM THICK PSILATE
	50    	AMPHIDIADEMA DENTICULATA
	51	AUSTRALISPHAERA SP
اا اا درون و من من من ا	52    	ISABELIDINIUM ROTUNDATA
	53    	ISABELIDINIUM TRIPARTITA
	54    	OLIGOSPHAERIDIUM PULCHERRIMUM
× · · · · × · · ·		
×	56	AEQUITRIRADITES TILCHAENSIS
× · · · · · × × · ·	57    	AEQUITRIRADITES VERRUCOSUS
111212 31	581	
X1XC1X141C	59    	
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× · · · · · · · · ·	63	j COPTOSPORA PARADOXA ! !
110	64	1
.3	65	CRYBELOSPORITES STRIATUS  -
25 11 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	66  	  L
7 7 111 13 4 4 16 9 9 11 111	67  	CYATHIDITES MINOR
× · · · · · · · · · ·	68  	ij CYCADOPITES FOLLICULARIS ∥
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₽···×.	70    	ii DICTYOTOSPORITES COMPLEX     -
$\times \cdot \cdot \cdot \cdot \cdot \cdot \cdot \cdot$	() 71    	
12 2 3 3 4 24 21 1	72   	DILWYNITES GRANULATUS
40821.1133	jj 73    	FALCISPORITES GRANDIS
1336611112	74    	FALCISPORITES SIMILIS
×·····	jj 75    	ij FORAMINISPORIS ASYMMETRICUS ‼ !
×·····	ji 76	

1388.0 SWC 3 1391.0 SWC 1500.0 SWC 1515.0 SWC 1650.0 SWC 1658.0 SWC 1672.0 SWC 1758.0 SWC 1763.0 SWC 1763.0 SWC	H H H H H H	
0    0    1	77    1	FORAMINISPORIS WONTHAGGIENSIS
2. 1. 1		KLUKISPORITES SCABERIS
H X X		LEPTOLEPIDITES VERRUCATUS
11 12 13 14 15 11 13	80    	MICROCACHRYIDITES ANTARCTICUS
Α	81	NEVESISPORITES VALLATUS
111134247.	82	OSMUNDACIDITES WELLMANII
σωω	83	PERINOPOLLENITES ELATOIDES
		PODOSPORITES MICROSACCATUS
" " + .4++ .	85	RETITRILETES AUSTROCLAVATIDITES
×	86 li	REWORKING - PERMIAN
83325731	87	STERIESPORITES ANTIQUASPORITES
× · · · · · · · · ·	88	TRIPOROLETES RETICULATUS
	89	VITREISPORITES PALLIBUS
. , <u>, , , , , , , , , , , , , , , , , ,</u>	90	CALLIALASPORITES TURBATUS
🗙 🗆 9 🗙 3 🗙 1	   91    	AMOSOPOLLIS CRUCIFORMIS
×	    92    	APPENDICISPORITES DISTOCARINATUS
× ×	    93    	BALMEISPORITES HOLODICTYUS
	    94    	DICTYOPHYLLIDITES
2122146	    95   	GLEICHENIIDITES
×	96       96    	INTERULOBITES INTRAVERRUCATUS
4. 1. 112	    97    	
H. NXXGB	    98	PHYLLOCLADIDITES MAUSONII
×	    99    	TRICOLPITES SPP
ω	100	TRICOLPITES VARIVERRUCATUS
	101	AUSTRALOPOLLIS OBSCURIS
× . ×	   102   	PEROTRILETES MAJUS 
	- 11	   TRICOLPITES GILLII 
×	   104 	   TRILOBOSPORITES TRIORETICULOSUS 
μμ ω	li	    CLAVIFERA TRIPLEX 
× . » .	11	    CAMEROZONOSPORITES OHAIENSIS 
×	   107 	    CONTIGNISPORITES COOKSONIAE 
4	   108	    PROTEACIDITES SPP 
× .	   109	    ORNAMENTIFERA SENTOSA 
	11	   PHIMOPOLLENITES PANNOSUS 
	ii.	TRICOLPORITES APOXYEXINUS
51 4 17 5 1 4 17	ii .	BOTRYOCACCUS
. ×	11	SCHIZOSPORIS PSILATUS
×	114 	SCHIZOSPORIS RETICULATUS

#### SPECIES LOCATION INDEX

Index numbers are the columns in which species appear.

43

INDEX NUMBER	SPECIES
1	MICROPLANKTON CONTENT (%)
55	AEQUITRIRADITES SPINULOSUS
56	AEQUITRIRADITES TILCHAENSIS
57 7	AEQUITRIRADITES VERRUCOSUS
91	ALTERBIA ACUMINATUM AMOSOPOLLIS CRUCIFORMIS
50	AMPHIDIADEMA DENTICULATA
92	APPENDICISPORITES DISTOCARINATUS
8	APTEA SP
58 51	ARAUCARIACITES AUSTRALIS
101	AUSTRALISPHAERA SP AUSTRALOPOLLIS OBSCURIS
93	BALMEISPORITES HOLODICTYUS
112	BOTRYOCOCCUS
25 90	CALLAOISPHAERIDIUM ASYMMETRICUM
106	CALLIALASPORITES TURBATUS CAMEROZONOSPORITES OHAIENSIS
45	CANNINGIA SPP
30	CASSICULOSPHAERIDIA GIANT
59	CERATOSPORITES EQUALIS
40 9	CHATANGIELLA VICTORIENSIS CHLAMYDOPHORELLA AMBIGUA
60	CICATRICOSISPORITES AUSTRALIENSIS
61	CICATRICOSISPORITES HUGHESI
10	CIRCULODINIUM DEFLANDREI
105 107	CLAVIFERA TRIPLEX
62	CONTIGNISPORITES COOKSONIAE CONTIGNISPORITES GLEBULENTUS
63	COPTOSPORA PARADOXA
64	COROLLINA TOROSA
26 11	CRIBROPERIDINIUM EDWARDSII
65	CRIBROPERIDINIUM SPP CRYBELOSPORITES STRIATUS
66	CYATHIDITES AUSTRALIS
67	CYATHIDITES MINOR
68	CYCADOPITES FOLLICULARIS
27 69	CYCLONEPHELIUM MEMBRANIPHORUM CYCLOSPORITES HUGHEST
94	DICTYOPHYLLIDITES
70	DICTYOTOSPORITES COMPLEX
71 72	DICTYOTOSPORITES SPECIOSUS
41	DILWYNITES GRANULATUS EUCLADINIUM MADURENSE
12	EXOCHOSPHAERIDIUM PHRAGMITES
73	FALCISPORITES GRANDIS
74 13	FALCISPORITES SIMILIS
75	FLORENTINIA DEANEI FORAMINISPORIS ASYMMETRICUS
76	FORAMINISPORIS DAILYI
77	FORAMINISPORIS WONTHAGGIENSIS
31 32	FROMEA FRAGILIS
95	GILLINIA HYMENOPHORA GLEICHENIIDITES
42	HETEROSPHAERIDIUM CF LATEROBRACHIUS
4	HETEROSPHAERIDIUM CONJUNCTUM
14 46	HETEROSPHAERIDIUM HETEROCANTHUM HETEROSPHAERIDIUM ROBUSTA
15	HETEROSPHAERIDIUM SOLIDA
28	HYSTRICHODINIUM PULCHRUM
96	INTERULOBITES INTRAVERRUCATUS
47 33	ISABELIDINIUM BELFASTENSE ISABELIDINIUM CRETACEUM
52	ISABELIDINIUM ROTUNDATA
29	ISABELIDINIUM SP
53	ISABELIDINIUM TRIPARTITA
16 17	KIOKANSIUM POLYPES KIOKANSIUM RECURVATUM
78	KLUKISPORITES SCABERIS
97	LAEVIGATOSPORITES OVATUS
79 2	LEPTOLEPIDITES VERRUCATUS
80	MICRHYSTRIDIUM MICROCACHRYIDITES ANTARCTICUS
81	NEVESISPORITES VALLATUS
18	NUMMUS SP
34 35	ODONTOCHITINA COSTATA ODONTOCHITINA CRIBROPODA
33 19	ODONTOCHITINA CRIBROPODA ODONTOCHITINA OPERCULATA
48	ODONTOCHITINA STUBBY
36	ODONTOCHITINIA PORIFERA

ODONTOCHITINIA PORIFERA

18	NEVESISPORITES VALLATUS
18	NUMMUS SP
34	ODONTOCHITINA COSTATA
35	ODONTOCHITINA COSTATA ODONTOCHITINA CRIBROPODA
19	ODONTOCHITINA CRIBROPODA ODONTOCHITINA OPERCULATA
48	ODONTOCHITINA STUBBY
36	ODONTOCHITINIA PORIFERA
43	ODONTOCHITINIA TRIANGULARIS
20	OLIGOSPHAERIDIUM COMPLEX
54	OLIGOSPHAERIDIUM PULCHERRIMUM
109	ORNAMENTIFERA SENTOSA
82	OSMUNDACIDITES WELLMANII
21	PALAEOHYSTRICHOSPHORA INFUSORIOIDES
37	PALAEOPERIDINIUM CRETACEUM
44	PARALECANIELLA
83	PERINOPOLLENITES ELATOIDES
102	PEROTRILETES MAJUS
110	PHIMOPOLLENITES PANNOSUS
98	PHYLLOCLADIDITES MAWSONII
84	PODOSPORITES MICROSACCATUS
108	PROTEACIDITES SPP
22	RECTANGULADINIUM SP
85	RETITRILETES AUSTROCLAVATIDITES
86	REWORKING - PERMIAN
113	SCHIZOSPORIS PSILATUS
114	
23	SPINIFERITES FURCATUS/RAMOSUS
87	STERIESPORITES ANTIQUASPORITES
5	SUBTILISPHAERA SP
24	TRICHODINIUM
103	TRICOLPITES GILLII
99	TRICOLPITES SPP
100	TRICOLPITES VARIVERRUCATUS
111	TRICOLPORITES APOXYEXINUS
104	TRILOBOSPORITES TRIORETICULOSUS
88	TRIPOROLETES RETICULATUS
6	TRITHYRODINIUM MARSHALLII
38	TRITHYRODINIUM PUNCTATE
49	TRITHYRODINIUM THICK PSILATE
39	TRITHYRODINIUM THICK RETICULATE
3	VERYHACHIUM
89	VITREISPORITES PALLIDUS

## APPENDIX 4

#### **APPENDIX**

4. Petrolab Compositional Analysis Report

47 Woodforde Road, Magill, South Australia, 5072 O. Box 410, agill, South Australia, 5072



Fax: 364 1500 Telex: AA88214 **Tel: (08) 364 1500** 

Reservoir Fluid and Core Services, Laboratory Consulting and Analysis

A. C. N. # 008 130 667

Adelaide, August 9, 1994 P.O. Box 410 Magill, S.A. 5072

Bridge Oil Limited #255 Elizabeth Street Sydney N.S.W. 2000 **Subject: Compositional Analysis** 

Well: Mylor #1 File: B-94045

Attention: Mr. Phillip Reichardt

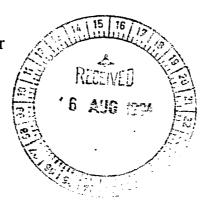
Dear Sirs,

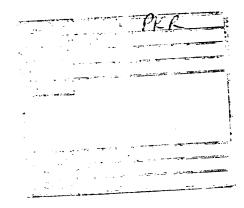
Please find enclosed results of compositional analyses of Mylor #1. The appearance of the liquid was very transparent, being practically colourless.

It has been a pleasure to have been of service. Please do not hesitate in contacting us should you require any further information or if we can assist you in any other way.

Yours Sincerely,

Marcel Volant Laboratory Manager





Company : Bridge Oil Limited Well : Mylor # 1

ll : Mylor # 1 File : B 94045

#### **REPORT INDEX**

Page

Summary Of Results	1
Transfer Details	2
Validity Check	3
Compositional Analyses of Equilibrated Gas	4
Compositional Analyses of Equilibrated Liquid	5-6
Constant Mass Study	7
·	
PLOTS:	
Relative Volume	8
Liquid Compressibility	9
Y—Function	10
Liquid Volume Factor	11
Liquid Viscosity at Reservoir Temperature	12

Company: Bridge Oil Limited

Well: Mylor # 1

Page: 2 of 12 File: B 94045

#### **SUMMARY OF RESULTS**

#### **CONSTANT MASS DATA:**

#### SATURATED LIQUID @ 150 °F AND 2775 PSIG:

Reservoir Temperature (°F) :	150
Saturation Pressure (psig) :	2775
Formation Volume Factor (Bbl/Bbl) :	2.1351
Liquid density (gm/cc) :	0.4997
Thermal Expansion @ 5000 psig (1/°F):	0.001464
(1/°C):	0.002635
Compressibility of saturated liquid	
@ 150 °F and 2775 psig (1/psi) :	42.50 x 10^-6
Viscosity (cp)	0.258

#### FLASH DATA:

#### SATURATED LIQUID @ 150 °F AND 2301 PSIG:

Solution GOR (SCF/bbl)	;	1406
Formation Volume Factor (Bbl/Bbl)	:	1.8662
Liquid density (gm/cc)	:	0.5403
Specific Volume (ft3/lb)	:	0.02965
Viscosity (cp)	:	0.210

#### **RESIDUAL LIQUID:**

API Gravity @ 60 °F	:	61.9
Density @ 150 °F (gm/cc)	:	0.6888
Viscosity @ 150 °F (cp)	:	0.572

Company: Bridge Oil Limited

Well: Mylor # 1

Page : 2 of 12

File: B 94045

#### TRANSFER DETAILS

#### **RUN #2**

Reservoir Pressure

2316 psia

Reservoir Temperature:

150 °F

RFT Chamber #

RFS # AE - 1238

Capacity

23/4 Gallon

Depth : Received :

1702.3 mKB July 8, 1994

**Opening Pressure** 

90 Psig @ 55 °F

Chamber compressed to 5000 psig with approximately 10480 cc"s of water behind the piston. Transferred 490 cc's into Petrolab cylinder L-104. Recovered another 2 cc's of water and 8 cc's of muddy filtrate. Chamber returned to Schlumberger Seaco Inc./GFR Resources, Victoria on July 13, 1994.

Company: Bridge Oil Limited Well: Mylor # 1

Page : 3 of 12 File : B 94045

# Bottom Hole Sample Validity Check

Sampling Conditions Sample # 1

2316 psia	150 °F	2	RFT Chamber #: RFS # AE - 1238	Capacity: 23/4 Gallon	Depth: 1702.3 mKB	90 psig @ 55 °F	L-104
Reservoir Pressure: 2316 psia	Reservoir Temperature: 150 °F	Run # : 2	RFT Chamber #:	Capacity:	Depth:	Opening Pressure: 90 psig @ 55 °F	Transferred into: L-104

Pressure (psig)	
Volume (cc's)	

2095	2285	2540	2735	2865	2895	3025	3295	3665	4076	4440	4965
30.00	40.00	50.00		58.00	59.00	60.50	62.00	64.00	90.99	68.00	70.00

(*)					I	
	11					 2
	14	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	Z X			so cted)
		-				Ha inje
				3		50 (cc's of
						VOLUME (cc's of Ha injected)
					₩ .	30
	4500	4000 1	0005		0000	 8
		4500		(bisq) =noce=1		3500 3500 3500 3500 3500 3500

Saturation Pressure : 2950 psig @ 61 ° F

Company: Bridge Oil Limited

Well: Mylor # 1

Page: 4 of 12

File: B 94045

#### COMPOSITIONAL ANALYSIS OF Gas Equilibrated @ 2301 psig and 150 ° F

From RFS # AE - 1238 Cylinder # L-104

Component		Mol %	GPM	
Hydrogen Sulphide	H2S	0.00		Pressure Base : 14.696 Zsc : 0.997
Carbon Dioxide	CO2	0.12		
Nitrogen	N2	3.32		Mol Weight : 19.77
Methane	C1	89.91		Gas Gravity : 0.684 Pc : 652.3
Ethane	C2	1.62	0.434	Tc : 369.1
Propane	C3	0.20	0.055	Mol Weight C6+ : 93.4
Iso-Butane	iC4	0.12	0.039	Density C6+ : 0.6800
N-Butane	nC4	1.19	0.376	Mol Weight C7+ : 100.3  Density C7+ : 0.6895
Iso-Pentane	iC5	0.69	0.253	Mol Weight C8+ : 112.6
N-Pentane	nC5	0.55	0.199	Density C8+ : 0.7048
				Mol Weight C11+:
Hexanes	<b>C6</b>	0.97	0.377	Density C11+ :
Heptanes	C7	0.97	0.408	Mol Weight C12+ : Density C12+ :
Octanes	C8	0.24	0.109	
Nonanes	C9	0.06	0.030	Heating Value (BTU/ft3)  Gross : 1157
Decanes	C10	0.04	0.022	Nett : 1048
Undecanes	C11	0.00	0.000	Wobbe Index : 1399
Dodecanes Plus	C12+	0.00	0.000	
TOTAL		100.00	2.302	Zpt * : 0.845

(P)ressure: 2301 psig (T)emperature: 150 °F

(G)radient: 0.05748 psi/feet

(19.77 \* 2315.7) / (0.845 \* 10.73 \* 610 \* 144)

Company: Bridge Oil Limited

Well: Mylor # 1

Page : 5 of 12 File : B 94045

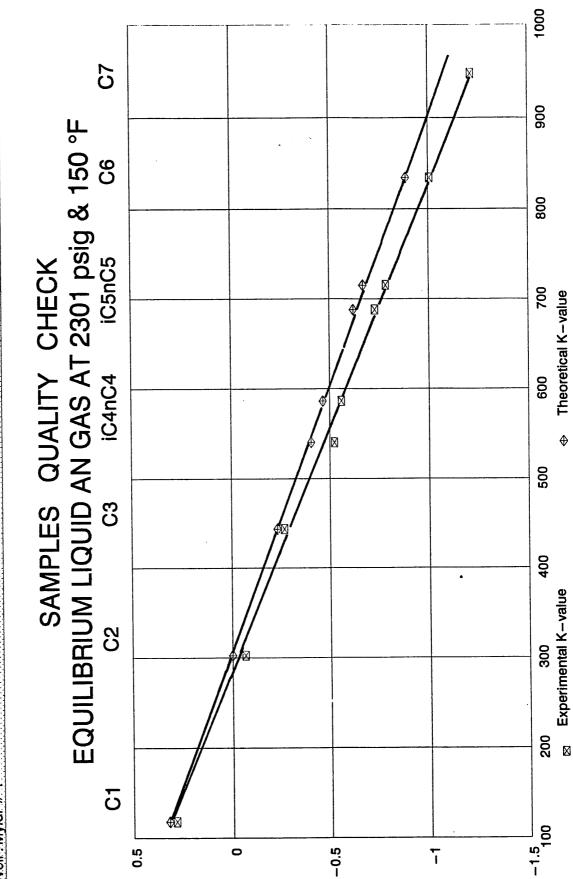
#### COMPOSITIONAL ANALYSIS OF STABILISED LIQUID AT 2316 psia and 150 deg F RFS # AE - 1238 Cyl. # L-104

	Stock Tank	Stock Tank	Stabilised
	Liquid -	Gas	Liquid
Component	Mol %	Mol %	Mol %
Hydrogen Sulphide	H2S 0.00	0.00	0.00
Carbon Dioxide	CO2 0.00	0.16	0.10
Nitrogen	N2 0.00	1.51	0.90
Methane	C1 0.47	78.26	46.62
Ethane	C2 0.12	3.10	1.89
Propane	C3 0.08	0.56	0.37
Iso-Butane	iC4 0.20	0.53	0.40
N-Butane	nC4 2.92	5.27	4.32
Iso-Pentane	iC5 4.66	3.07	3.72
N-Pentane	nC5 4.77	2.41	3.37
Hexanes	C6 19.82	2.99	9.83
	C7 36.47	1.87	15.94
Heptanes	C8 10.63	0.25	4.47
Octanes			
Nonanes	C9 8.75	0.02	3.57
Decanes	C10 4.98	0.00	2.03
Undecanes	C11 2.36	0.00	0.96
Dodecanes Plus	C12+ 3.76	0.00	1.51
TOTAL	100.00	100.00	100.00
Ratios			
Molar Ratio	: 0.4068	0.5932	1.0000
Mass Ratio Liquid Ratio (bbl/bbl)	: 0.7233 : 1.0000 @ SC	0.2767	1.0000 1.8520 @ PT*
Gas Liquid Ratio	: 1.0000 bbl @ SC	1406 SCF	
Stream Properties			
Molecular Weight	: 99.7	26.15	54.1
Density obs. (gm/cc) Gravity (AIR = 1.000)	: 0.7225 @ 60 °F : 64.2 °API @ 60 °F	0.908	0.5403 @ PT*
GHV (BTU/scf)	:	1533.0	
Hexanes Plus Prope	rties		
Mol %	: 86.76	5.13	28.48
Molecular Weight	: 104.8	69.6	103.6
Density (gm/cc @ 60 °F) Gravity (°API @ 60 °F)	: 0.7357 : 60.6	0.6746 78.0	0.7312
Glavity (AFT @ GO F)			618 1
		76.0	61.8
Heptanes Plus Prop			
Mol %	: 66.94	2.14	12.54
Mol % Molecular Weight			
Mol %	: 66.94 : 85.6	2.14 40.7	12.54 83.6
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity (°API @ 60 °F) Decanes Plus Prope	: 66.94 : 85.6 : 0.7481 : 57.5	2.14 40.7 0.5702 116.4	12.54 83.6 0.7431 58.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol %	: 66.94 : 85.6 : 0.7481 : 57.5	2.14 40.7 0.5702 116.4	12.54 83.6 0.7431 58.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9	2.14 40.7 0.5702 116.4	12.54 83.6 0.7431 58.7 4.50 155.9
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol %	: 66.94 : 85.6 : 0.7481 : 57.5	2.14 40.7 0.5702 116.4	12.54 83.6 0.7431 58.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F) Undecanes Plus Pro	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7	2.14 40.7 0.5702 116.4 0.00	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity (°API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity (°API @ 60 °F)  Undecanes Plus Pro Mol %	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7	2.14 40.7 0.5702 116.4 0.00   	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7  perties : 6.12 : 173.8	2.14 40.7 0.5702 116.4 0.00 	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol %	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7	2.14 40.7 0.5702 116.4 0.00   	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density ("API @ 60 °F)	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7  perties : 6.12 : 173.8 : 0.8109 : 42.8	2.14 40.7 0.5702 116.4 0.00     0.00	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7 2.47 173.8 0.8109 42.8
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Cravity ("API @ 60 °F)  Dodecanes Plus Pro Mol %	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7  perties : 6.12 : 173.8 : 0.8109 : 42.8	2.14 40.7 0.5702 116.4 0.00    0.00    0.00	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7 2.47 173.8 0.8109 42.8
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density (gm/cc @ 60 °F)  Dodecanes Plus Pro Mol % Molecular Weight	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7  perties : 6.12 : 173.8 : 0.8109 : 42.8  perties : 3.76 : 190.7	2.14 40.7 0.5702 116.4 0.00    0.00    0.00	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7 2.47 173.8 0.8109 42.8
Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Decanes Plus Prope Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)  Undecanes Plus Pro Mol % Molecular Weight Density (gm/cc @ 60 °F) Gravity ("API @ 60 °F)	: 66.94 : 85.6 : 0.7481 : 57.5  rties : 11.10 : 155.9 : 0.7979 : 45.7  perties : 6.12 : 173.8 : 0.8109 : 42.8	2.14 40.7 0.5702 116.4 0.00    0.00    0.00	12.54 83.6 0.7431 58.7 4.50 155.9 0.7979 45.7 2.47 173.8 0.8109 42.8

<sup>\* (</sup>P)ressure: 2301 psig

Page : 6 of 12 File : B 94045

Company : Bridge Oil Limited Well : Mylor # 1



LOG EQUILIBRIUM RATIO

(TC) ~ 2 OF PARAFFIN COMPONENT (/1000)

Company: Bridge Oil Limited Page: 7 of 12
Well: Mylor # 1 File: B 94045

### CONSTANT MASS STUDY @ 150 ° F

Pressure (psig)	Relative Volume (V/Vsat)	Liquid Compressibility (x10 ^ -6)(psig ^ -1)	Y Function	Formation Volume Factor (Bo)	Liquid Viscosity (cp)
	(1)	(2)	(3)	(4)	
5000	0.9434	18.50		2.0143	0.258
4552	0.9517	20.65		2.0321	0.242
4086	0.9615	23.25		2.0529	0.229
3669	0.9715	26.32		2.0743	0.215
3254	0.9831	30.75		2.0990	0.203
3047	0.9897	34.15		2.1131	0.195
2832	0.9976	40.15		2.1301	0.190
2775 *	1.0000	42.50		2.1351	0.189
2685	1.0161		2.350	2.0869	0.191
2559	1.0427		1.978	1.9758	0.197
2435	1.0731		1.826	1.9098	0.201
2301 **	1.1104		1.739	1.8662	0.210
2229	1.1332		1.711	1.8385	0.211
2026	1.2077		1.625	1.7798	0.222
1862	1.2820		1.572	1.7390	0.230
1607	1.4352		1.505	1.6702	0.245
1416	1.5893		1.474	1.6286	0.260
1262	1.7522		1.450	1.5938	0.270
1018	2.1165		1.402	1.5459	0.290
897	2.3769		1.397	1.5206	0.300
700	3.0081		1.358	1.4762	0.321
630	3.3355		1.348	1.4578	0.330
0				1.0613	0.572

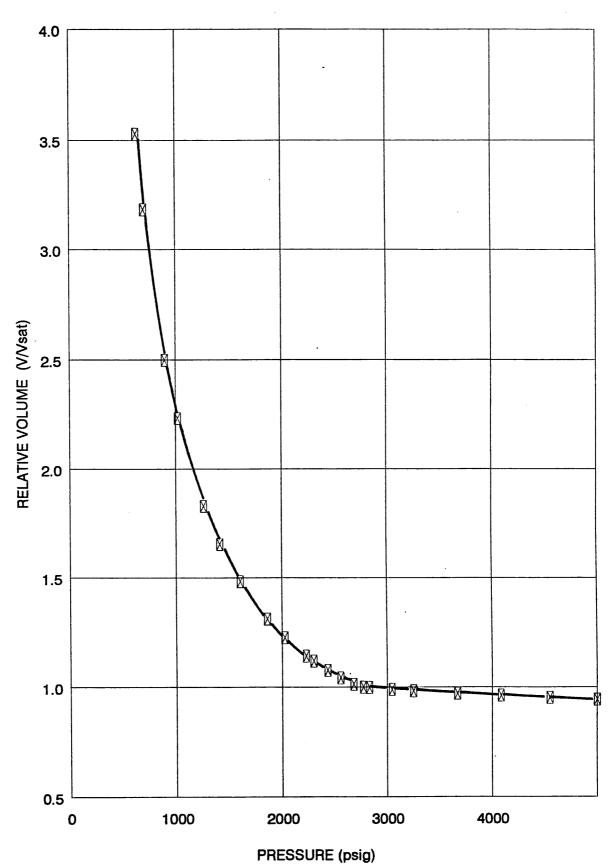
- Saturation pressure
- \*\* Reservoir pressure
- (1) Barrels at indicated pressure per barrel at saturation pressure
- (2) Liquid Compressibility = -(1/V) \* (dV/dP)
- (3) Y Function = (Psat P) / (P)\*(V/Vsat-1)
- (4) Barrels of liquid at indicated pressure and temperature per barrel of residual liquid at 60 °F.

Company: Bridge Oil Limited Well: Mylor # 1

Page: 8 of 12

File: B 94045

#### RELATIVE VOLUME



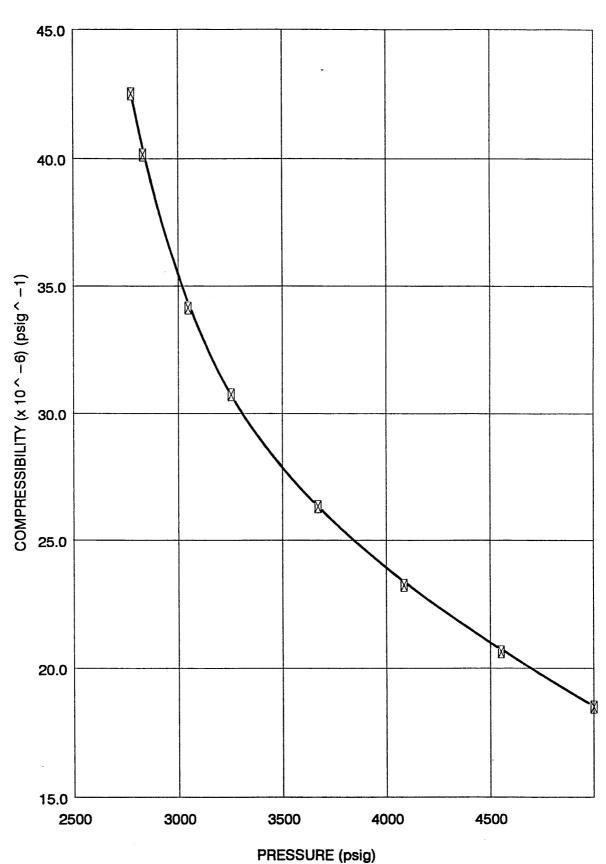
Company: Bridge Oil Limited

Well: Mylor # 1

Page: 9 of 12

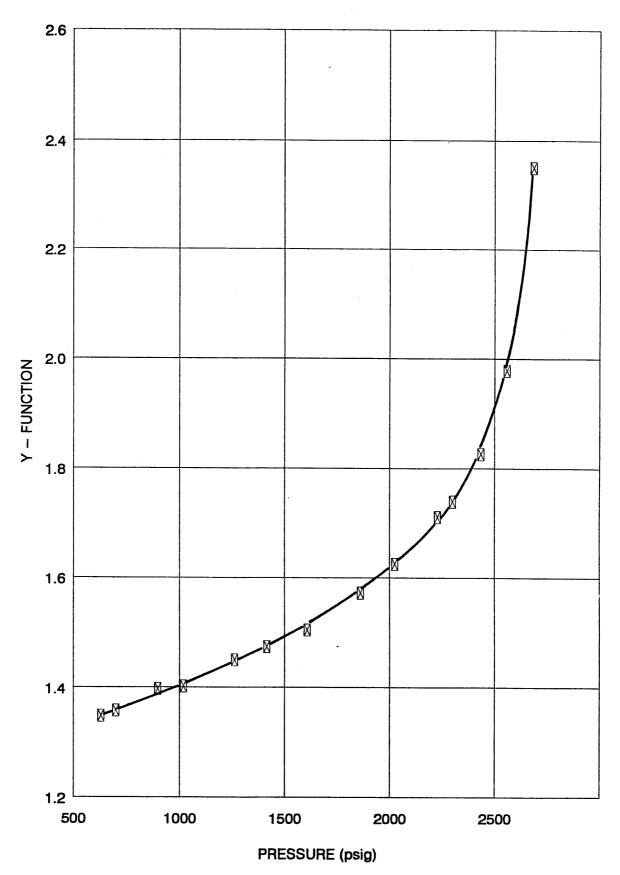
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#### LIQUID COMPRESSIBILITY



Company: Bridge Oil Limited Page: 10 of 12
Well: Mylor # 1 File: B 94045

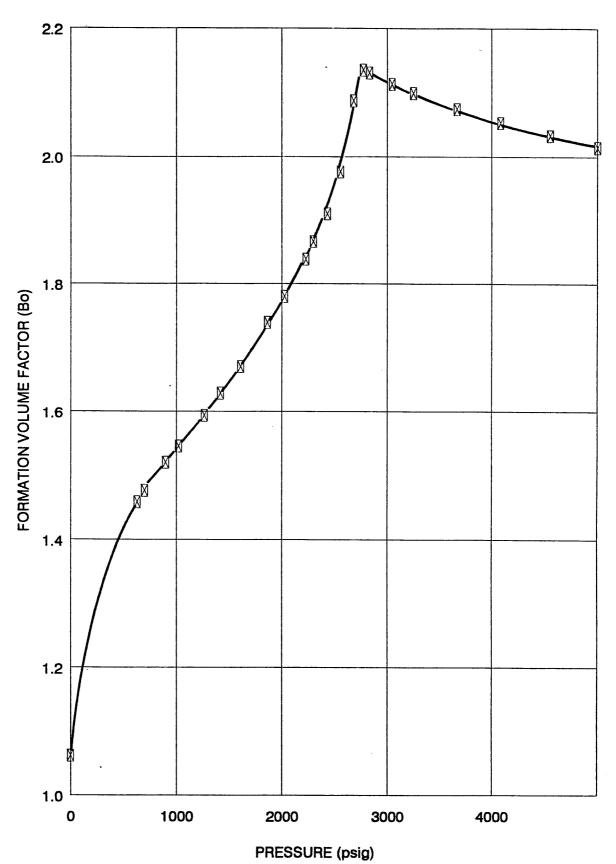
#### Y - FUNCTION



Company: Bridge Oil Limited
Well: Mylor # 1

Page: 11 of 12 File: B 94045

#### LIQUID VOLUME FACTOR



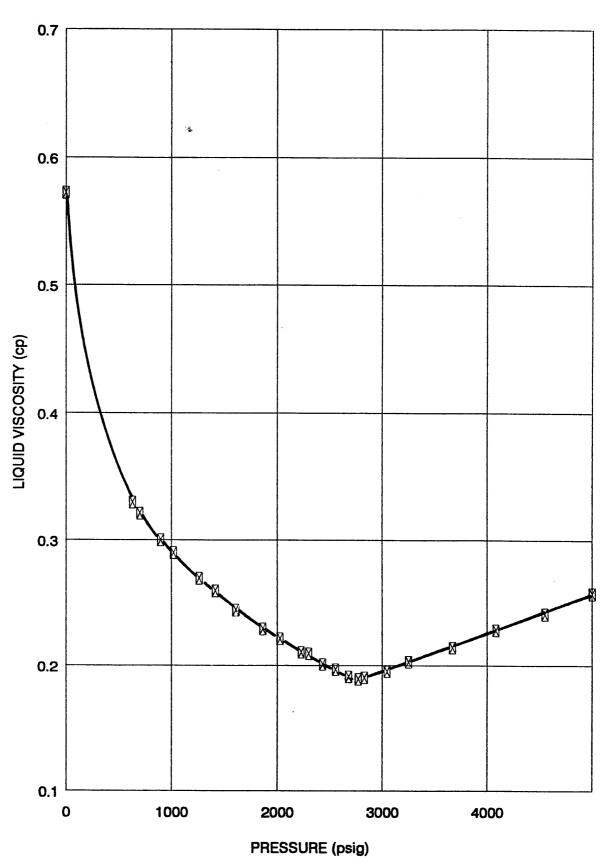
Company: Bridge Oil Limited

Well: Mylor # 1

Page: 12 of 12

File: B 94045

#### LIQUID VISCOSITY



# APPENDIX 5

### **APPENDIX**

5. Amdel Geochemical Evaluation of Core, Cuttings and Condensate Samples from Mylor #1



### GEOCHEMICAL EVALUATION OF CORE, CUTTINGS AND CONDENSATE SAMPLES FROM

**MYLOR-1** 

**OTWAY BASIN, VICTORIA** 

REPORT LQ3108 FOR

**BRIDGE OIL LIMITED** 

BY
SCOTT WYTHE
BRIAN WATSON



18 October 1994

Amdel Limited A.C.N. 008 127 802

Petroleum Services PO Box 338 Torrensville SA 5031

Telephone: (08) 379 9888 Facsimile: (08) 379 6623

Bridge Oil Limited PO Box A1195 SYDNEY SOUTH NSW 2000

Attention: Barry Goldstein

### **REPORT LO3108**

**CLIENT REFERENCE:** 

PO 4254 & 7396

WELL NAMES:

Mylor-1

**MATERIAL:** 

Core, Cuttings and Condensate samples

**WORK REOUIRED:** 

Source Rock and Oil Geochemistry

Please direct technical enquiries regarding this work to the signatory below under whose supervision the work was carried out.

Brian L. Watson

Brin Water

Manager

**Petroleum Services** 

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### **CONTENTS**

2.	ANAI	ANALYTICAL PROCEDURES					
3.	RESU	JLTS					
4.	INTE	INTERPRETATION					
	4.1	Source Rock Geochemistry					
	4.1.2	Maturity Source Richness Kerogen Type and Source Quality					
	4.2	Oil Geochemistry					
		Maturity Source Affinity					
	4.3	Post Pooling Alteration and Migration					
5.	CON	CLUSIONS					

**INTRODUCTION** 

**1.** 

### LIST OF TABLES

.1.	Vitrinite Reflectance Data
2.	Maceral Group Proportions
3.	Organic Matter Type and Abundance
4.	Liptinite Maceral Abundance and Fluorescence Characteristics
5.	Rock-Eval Pyrolysis Data
6.	Gasoline Range Analysis
7.	Extract Yields and Alkane Ratios
8.	Saturated Biomarker Ratios
9.	Oil Maturity Based on Aromatic Hydrocarbon Distributions

### LIST OF FIGURES

1.	Vitrinite Reflectance versus Depth
2.	Hydrogen Index versus T <sub>max</sub>
3.	Mylor-1 DST-1 Condensate, Gasoline Range Chromatogram
4.	Oil Maturity and Alteration based on C <sub>5</sub> -C <sub>7</sub> alkanes
5.	Oil Source Affinity based on C <sub>5</sub> -C <sub>7</sub> alkanes
6.	Mylor-1 1685.74-1685.86 metres, GC of Extractable Organic Matter
7.	Mylor-1 1702.9 metres, GC of Extractable Organic Matter
8.	Mylor-1 DST-1 Condensate, GC of Saturates Fraction
9.	Genetic Affinity based on Isoprenoid/Alkane Ratios
10.	Oil Source Affinity Based on Saturated Biomarker GC-MS Data
11.	Sterane Maturity - Migration Plot
12.	Sterane Distributions
13.	Araucariacean Resin Biomarker Plot

### LIST OF APPENDICES

.1.	Analytical Procedures
2.	Vitrinite Reflectance Values and Histograms
3.	Photomicrographs
4.	GC-MS of Branched/Cyclic Alkanes
5.	GC-MS of Aromatic Hydrocarbons

### 1. INTRODUCTION

Three (3) cuttings samples and four (4) sidewall core samples were received from Mylor-1 for TOC analysis and Rock-Eval Pyrolysis along with vitrinite reflectance and organic petrological analysis. In addition two (2) core samples and one (1) DST-1 (1665.7 - 1684m KBD) condensate sample from the same well were received for oil geochemical analysis. An RFT oil was also submitted by Dr.D.M.McKirdy for gasoline range analysis and gas chromatography. Results of analyses on this RFT sample have been reported to Dr. McKirdy and will only be discussed briefly here. This report is a formal presentation of results forwarded as they became available.

### 2. ANALYTICAL PROCEDURES

The analytical procedures used in this study are provided in Appendix 1.

### 3. RESULTS

Analytical data is presented in this report as follows:

Analysis	Table	Figure
Vitrinite Reflectance Analysis and Organic Petrology	1-4	1 Appendices 2 & 3
Rock-Eval Pyrolysis	5	2
Gasoline Range Analysis	6	3 - 5
GCs of Extracted Hydrocarbons	7	6 - 9
GC-MS of branched/cyclic hydrocarbons	8	10 - 12 Appendix 4
GC-MS of aromatic hydrocarbons	9	13 Appendix 5

### 4. INTERPRETATION

### 4.1 Source Rock Geochemistry

### 4.1.1 Maturity

Measured vitrinite reflectance values range from 0.40-0.53% in the samples analysed (Table 1). Figure 1 indicates that the sedimentary section intersected in Mylor-1 should be sufficiently mature for the generation of light oil/condensate from thermally labile liptinites (resinite, suberinite and bituminite) below approximately 1300 metres (VR = 0.45%). The maturity threshold for significant gas generation from terrestrial woody herbaceous kerogen (VR = 0.55%) occurs at approximately 2500 metres in this location. Below approximately 4000 metres the sedimentary interval is mature for the generation of liquid hydrocarbons from the less thermally labile liptinites (VR > 0.7%).

Rock-Eval  $T_{max}$  values and Hydrogen Indices (Table 5, Figure 2) suggest that the samples are immature to marginally mature (VR<sub>equiv</sub>=0.4-0.5%) which is in agreement with the measured vitrinite reflectance data.

### 4.1.2 Source Richness

Organic richness ranges from poor to fair in the samples studied (TOC = 0.14 - 3.06%; Table 5). Samples with fair organic richness occur in the interval 1386 - 1839 metres depth.

Source richness for the generation of hydrocarbons is generally poor  $(S_1 + S_2 = 0.33 - 0.60 \text{ kg of hydrocarbons/tonne}; Table 5)$  with the exception of the SWC from 1744 metres depth which has fair source richness  $(S_1 + S_2 = 2.36 \text{ kg of hydrocarbons/tonne}; Table 5)$ .

The likely presence of migrated oil is indicated by the high Production Index (PI = 0.37) in the sample from 1386 - 1389 metres depth.

### 4.1.3 Kerogen Type and Source Quality

Hydrogen Index and  $T_{max}$  values for the five samples plotted in Figure 2 indicate that these sediments contain organic matter which has bulk compositions ranging from Type III to Type IV kerogen.

Maceral group proportions derived from organic petrology analyses on these samples (Table 2) are consistent with the bulk compositions indicated by the Rock-Eval analyses. Samples consist largely of inertinite and have low to moderate vitrinite and liptinite contents (up to 15%).

Liptinite contents of up to 10% suggest the possibility of minor oil generation from samples from 1744 and 660 metres depth on maturity.

These indications would be more definitively assessed by pyrolysis - GC analyses.

The liptinite macerals present suggest a terrestrial environment of deposition. Lamalginite and phytoplankton are indicative of lacustrine facies (Tables 3 and 4).

### 4.2 Oil Geochemistry

### 4.2.1 Maturity

Aromatic maturity indicators for the DST-1 condensate (1665.7 - 1684m KBD) indicate that the sample was generated and expulsed from a mature source interval (Parameters A, C, E and F, Table 9). The equivalent vitrinite reflectance value of 0.83% (Parameter E, Table 5) is significantly higher than the measured vitrinite reflectance value at this depth (VR  $\approx 0.5\%$ , Table 1) indicating that the condensate has migrated some distance from depth.

Maturities indicated for this sample by saturated biomarker maturity parameters are less precise than the aromatic derived biomarker ratios. These parameters (ie,  $C_{29}$  steranes and isosteranes - Biomarker Parameters 4 & 6,  $C_{27}$  diasteranes - Biomarker Parameter 5,  $C_{27}$ ,  $C_{30}$  &  $C_{32}$  hopanes - Biomarker Parameters 9 - 11,  $C_{30}$  moretanes - Biomarker Parameter 12) generally support a moderate maturity for the condensate.

The two core extracts are from core just below the interval tested in DST-1 and along with the RFT oil have isoprenoid/n-alkane ratios (Table 7, Figure 9) very similar to that of the DST-1 condensate. This implies a similar maturity for all four of the samples.

### 4.2.2 Source Affinity

Pristane/phytane ratios (Table 7, Figure 10) for all four of the samples studied are indicative of generation from sources deposited in highly oxic conditions. Isoprenoid/n-alkane ratios suggest a similar higher plant source for the two extracts, the DST-1 condensate and the RFT sample.

Gasoline range analysis showed that the  $C_5$ - $C_7$  fractions of both the DST-1 condensate and the RFT sample have relatively high proportions of cyclic compounds which are most abundant in land plant derived organic matter. Differences between the gasoline range ratios for these two samples is likely to be due to more significant light end loss in the DST-1 sample.

GC-MS of branched/cyclic alkanes for the condensate has sterane and diasterane distributions (m/z 217, 218, 259; Table 8, Figures 10 & 12, Appendix 4) which are influenced by both  $C_{27}$  homologues of algal/bacterial origin and  $C_{29}$  homologues of higher plant origin (Biomarker parameters 1-3, Table 8). This suggests that the extracts contain either a mix of oils of different origin or an oil derived from a mixed source. The sample may therefore have been deposited in a near shore environment with both marine and terrestrial input.

Diterpanes and labdanes (m/z 123) are derived from sources containing resinite. Resinite is formed from higher plant resins and its presence implies that the source contains a component of terrestrial kerogen. These compounds are present in the DST-1 condensate sample.

Tricyclic and tetracyclic terpane compounds (m/z 191) are reasonably abundent in the condensate.  $C_{19}$  and  $C_{20}$  tricyclics which may be evidence of a higher plant source are present though not abundant in the sample. A substantial amount of the higher plant marker  $C_{24}$  tetracyclic terpane is however present in the sample.

The ratios of 1-methylphenanthrene/9-methylphenanthrene and 1,2,5-trimethylnaphthalene/1,3,6-trimethylnaphthalene (Figure 13) has been used to indicate source input from Araucariacean derived plant resins (trees from the Kauri pine group) which were most prominent in Early to Middle Jurassic times. The lack of Araucariacean source input of these compounds implies that these resins were not significant components of the precursor organic matter of the DST-1 condensate. However, this does not preclude the possibility that this sample was generated from a source of Jurassic/Cretaceous age.

### 4.3 Post Pooling Alteration and Migration

No evidence of significant biodegradation was seen for any of the samples analysed however isoprenoid/n-alkane ratios (Table 7, Figure 9) and gasoline range heptane values (Table 6, Figure 4) suggest that both the DST and RFT samples along with possibly the extracts may have been subjected to minor biodegradation. The very low aromatic contents of the DST-1 and RFT samples indicate that both these samples have undergone significant water washing.

Figure 13 along with differences between the aromatic maturity determination and the maturity of local sediments indicate that the DST-1 condensate is likely to have undergone significant migration since generation from its source interval(s).

### 5. CONCLUSIONS

- 5.1 Measured vitrinite reflectance values range from 0.40-0.53% in the samples analysed. Rock-Eval  $T_{max}$  values and Hydrogen Indices are in agreement with the vitrinite reflectance data.
- 5.2 Organic richness ranges from poor to fair in the samples studied with the samples in the interval 1386 1839 metres depth having fair organic richness. Source richness for the generation of hydrocarbons however is poor with the exception of the SWC from 1744 metres depth which has fair source richness.
- 5.3 Samples examined contain organic matter which has bulk compositions ranging from Type III to Type IV kerogen.
- 5.4 Maceral analyses showed liptinite contents of up to 10% suggesting minor oil generative potential. Liptinite macerals suggest a terrestrial environment with some lacustrine facies.
- 5.5 Aromatic maturity indicators for the DST-1 condensate indicate that the sample was generated and expulsed from mature source interval(s) ( $VR_{equiv} = 0.83\%$ ). This value is significantly higher than the measured vitrinite reflectance value at this depth (VR = 0.50%) indicating that the oil has migrated some distance from depth. Saturated biomarker maturity indicators and alkane ratios support a similar maturity level for this sample.
- Various aspects of the molecular composition of the DST-1 condensate indicates significant input of both higher plant and algal derived kerogen into the precursor organic matter suggesting the possibility of a near shore environment of deposition.
- 5.7 All of the samples appear to have been subjected to significant water washing and possibly a minor amount of biodegradation also.

TABLE 1
SUMMARY OF VITRINITE REFLECTANCE MEASUREMENTS, MYLOR-1

Depth (m)	Mean Maximum Reflectance (%)	Standard Deviation	Range	Number of Determinations
650-660	0.40	0.01	0.39-0.40	2
840-850	0.42	0.02	0.40-0.44	3
1386-1389	0.48	0.02	0.43-0.52	20
1550	0.53	0.02	0.51-0.57	6
1744	0.47	0.04	0.40-0.52	16
1835	0.50	0.04	0.43-0.57	18
1901	0.53	0.01	0.52-0.55	6

MACERAL GROUP PROPORTIONS

Depth	Percentage of					
( <b>m</b> )	Vitrinite Inertinite		Liptinite			
650-660	5	85	10			
840-850	<5	>90	<5			
1386-1389	5-10	90	<5			
1550	<5	90	5			
1744	10-15	80	5-10			
1835	<5	90	5			
1901	<5	90-95	<5			

ORGANIC MATTER TYPE AND ABUNDANCE

Depth	Relative	Estimated	Volume of	Exinite Macerals
( <b>m</b> )	(m) Maceral Group Proportions		Liptinite	
650-660	I>>L>V	~0.5	Ra-Vr	Bmite, Res, Lama
840-850	I>>V~L	< 0.5	Ra-Vr	Lipto
1386-1389	I>>V>L	1-2	Ra-Vr	Spo
1550	I>>L>V	~1	Ra	Lipto, Cut, Res, Phyto
1744	I>>V>L	2-3	Ra	Cut, Lama, Lipto, Spo, Res, Phyto
1835	I>>L>V	0.5-1	Ra-Vr	Lama, Lipto, Phyto
1907	I>>V~L	~0.5	Vr	Lipto

LIPTINITE MACERAL ABUNDANCE AND FLUORESCENCE CHARACTERISTICS

Depth (m)	Liptinite Macerals	Lithology/Comments
650-660*	Bmite (Ra; dO) Res (Vr; mY-mO), Lama (Vr; mO)	Cheifly sandstone with minor shale and carbonaceous shale.
840-850*	Lipto (Ra-Vr; mO)	Chiefly sandstone with minor shale.
1386-1389*	Spo (Ra-Vr; mO)	Chiefly shale with minor siltstone.
1550*	Lipto (Ra; mY-mO), Cut (Vr; mO), Res (Vr; iY), Phyto (Tr; mY)	Chiefly shale.
1744*	Cut (Ra; mO), Lama (Ra-Vr; mY-mO), Lipto (Ra-Vr; mO) Spo (Vr; mO), Res (Vr, mY), Phyto (Tr; mO)	Chiefly shale.
1835*	Lama (Ra-Vr; mO), Lipto (Ra-Vr; mO), Phyto (Vr; ?mY).	Chiefly shale.
1901*	Lipto (Vr; mO)	Cheifly siltstone.

<sup>\*</sup> Liptinite macerals are generally slightly to moderately oxidised.

TABLE 5

\*Estimated hand measured value

### **AMDEL PETROLEUM SERVICES**

			Re	ock – Ev	al Pyrolys	is				3	0/08/94
Client:	Bridge Oil Li	imited									
Well:	Mylor-1										
Depth (m)	Tmax	<b>S1</b>	<b>S2</b>	S3	S1+S2	PI	S2/S3	PC	TOC	н	OI
650660 840850									0.14 0.20		
1386-1389	420*	0.12	0.21	1.58	0.33	0.37	0.13	0.02	1.53	13	103
1550 SWC 25	430	0.05	0.47	1.22	0.52	0.10	0.39	0.04	1.45	32	84
1744 SWC 12	2 434	0.20	2.16	2.11	2.36	0.08	1.02	0.19	3.06	70	69
1839 SWC 4 1901 SWC 3	440*	0.06	0.54	0.31	0.60	0.10	1.74	0.05	0.61 0.29	88	51



### PETROLEUM SERVICES GASOLINE RANGE ANALYSIS

JOB NO:

LQ3108

**CLIENT:** 

Bridge Oil Limited

**WELL:** 

Mylor-1 DST-1 Condensate

Compound	Normal (%)	Branched (%)	Cyclic (%)	Aromatic (%)
2-Methylbutane		0.19		
n-Pentane	0.56			
2,2-Dimethylbutane		0.16		
Cyclopentane			0.00	
2,3—Dimethylbutane		0.59		
2-Methylpentane		2.36		
3-Methylpentane		1.62		
n-Hexane	6.37			
2,2—Dimethylpentane		0.61		
Methylcyclopentane			4.18	
2,4—Dimethylpentane		0.97		
2,2,3—Trimethylbutane		0.22		
Benzene				0.00
3,3—Dimethylpentane		0.45		0.00
Cyclohexane			9.21	
2-Methylhexane		5.02		
2,3-Dimethylpentane		1.80		
1,1—Dimethylcyclopentane			1.03	
3-Methylhexane		4.85		
Trans-1,3-Dimethylcyclopentane			1.68	
Cis-1,3-Dimethylcyclopentane			1.87	
3-Ethylpentane		0.00		
Trans-1,2-Dimethylcyclopentane			2.68	
n-Heptane	17.24			
Methylcyclohexane			35.76	·
Ethylcyclopentane			0.50	
Toluene				0.08
Total Percentages	24.17	18.84	56.91	0.08

Gasoline Range Parameters

	Gasonne Range Parameters					
Parameter	Specificity	Value				
n-hexane/methylcyclopentane	maturity/biodegradation	1.52				
n-heptane/methylcyclohexane	maturity/biodegradation	0.48				
3-methylpentane/benzene	water washing	0.00				
cyclohexane/benzene	water washing	0.00				
methylcyclohexane/toluene	water washing	436.90				
isopentane/normal pentane	maturity/biodegradation	0.34				
3-methylpentane/n-hexane	biodegradation	0.26				
isoheptane value *	maturity	1.58				
heptane value *	maturity	21.24				

(\* from Thompson, 1983)

TABLE 7

 $C_{12+}$  BULK COMPOSITION AND ALKANE RATIOS, MYLOR-1

Sample	Extract		Alkane	Alkane Ratios	
Depth (m)	Yield (ppm)	Np/Pr	hq/rq	Pr/n-C <sub>17</sub>	Ph/n-C <sub>18</sub>
1685.74-1685.86	71.93	0.44	6.3	0.44	0.08
1702.9	1429	0.42	6.2	0.47	0.08
DST-1	ı	0.50	8.4	0.41	0.08
RFT	ı	0.42	7.4	0.41	0.09

Np = norpristane
Pr = pristane
Ph = phytane

 $n-C_{17}$  = n-heptadecane  $n-C_{18}$  = n-octadecane

# BIOMARKER PARAMETERS OF SOURCE, MATURITY, MIGRATION AND BIODEGRADATION, MYLOR-1, DST-1

		Ste	Steranes						Ţ	Terpanes	7,5			Acyc	Acyclic Alkanes	anes
-							Para	Parameter							:	-
-	2	3	4	S	9	4	8	6	10	11	12	13 14	14	15	16	17
38:21:41	1.08	1.08   1.23	1.16   1.61	1.61	1.59	1.59 0.51	pu	pu	0.56	0.46	1.54	0.10	pu	8.4	nd 0.56 0.46 1.54 0.10 nd 8.4 0.41 0.08	0.08

# nd = not determined

# KEY TO BIOMARKER PARAMETERS OF SOURCE, MIGRATION AND BIODEGRADATION

Parameter	Derivation*	Specificity
1	$_{\rm 20}$ $5a({\rm H})14a({\rm H})$	Source
7	$C_{\infty}$ 5a(H)14a(H)17a(H) 20R sterane/ $C_{21}$ 5a(H)14a(H)1/a(H) 20R sterane	Source
m	$C_{\infty}$ 138(H)17 $\alpha$ (H) 20R diasterane/ $C_{\gamma}$ 136(H)17 $\alpha$ (H) 20R diasterane	Source
4	$C_{\infty} 5a(H)14a(H)17a(H)$ 20S sterane/ $C_{\infty} 5a(H)14a(H)17a(H)$ 20R sterane	Maturity, Biodegradation
2	$C_n 13b(H)17a(H)$ 208 diasterane/ $C_n 13b(H)17a(H)$ 20R diasterane	Maturity
9	$C_{\infty}^{2} 5a(H)148(H)176(H)$ 20R sterane/ $C_{\infty} 5a(H)14a(H)17a(H)$ 20R sterane	Maturity, Migration
7	$C_{29} 13\dot{a}(H)17\dot{a}(H) 20R + 20S$ diasteranes/ $C_{29} 5\dot{a}(H)$ steranes	Migration, Source
∞	18α(H)-30-norneohopane (C <sub>2</sub> Ts)/C <sub>2</sub> 17α(H) hopane + C <sub>2</sub> Ts	Maturity, Source
6	$17a(H)$ diahopane/ $18a(H)$ -30-norneohopane ( $C_{30}^*/C_{20}TS$ )	Source, Maturity
10	,30-tris	Maturity, Source
11	$T_{\rm e}/C_{\rm in}$ 17a(H)21a(H) hopane	Maturity
12	17a(H)21s(H) 22S	Maturity
13	$C_{30}^{-}$ 178(H)21 $\alpha$ (H) moretane/ $C_{30}$ 17 $\alpha$ (H)218(H) hopane	Maturity
14	17a(H)-25-norhopa	Biodegradation
15	pristane/phytane	Source
16	pristane/n-heptadecane	Source, Biodegradation, Maturity
17	phytane/n-octadecane	Source, Biodegradation, Maturity

\* Ratios calculated from peak areas as follows:

Parameters 1-7 m/z = 217, 218, 259 mass fragmentograms

Parameters 8 - 14 m/z = 191 mass fragmentogram

Parameters 15 - 17 capillary gas chromatogram of alkanes or whole oil/extract

TABLE 9

### AROMATIC MATURITY DATA, MYLOR-1 DST-1 CONDENSATE

						VR CA	LC (%)		
MPI	MPR	DNR	MPDF	A	В	C	D	E	F
0.874	1.300	5.468	0.512	0.92	1.78	1.05	3.41	0.83	0.98

### KEY TO AROMATIC MATURITY INDICATORS

Methylphenanthrene index (MPI), methylphenanthrene ratio (MPR), dimethylnaphthalene ratio (DNR) and calculated vitrinite reflectance (VR<sub>calc</sub>) are derived from the following equations (after Radke and Welte, 1983; Radke *et al.*, (1984):

$$MPI = \frac{1.5(2\text{-MP} + 3\text{-MP})}{P + 1\text{-MP} + 9\text{-MP}}$$

$$VR_{calc} (a) = 0.6 \text{ MPI} + 0.4 \text{ (for VR} < 1.35\%)$$

$$VR_{calc} (b) = -0.6 \text{ MPI} + 2.3 \text{ (for VR} > 1.35\%)$$

$$MPR = \frac{2\text{-MP}}{1\text{-MP}}$$

$$VR_{calc} (c) = 0.99 \log_{10} \text{ MPR} + 0.94 \text{(VR} = 0.5\text{-}1.7\%)$$

$$DNR = \frac{2.6\text{-DMN} + 2.7\text{-DMN}}{1.5\text{-DMN}}$$

$$VR_{calc} (d) = 0.46 \text{ DNR} + 0.89 \text{ (for VR} = 0.9\text{-}1.5\%)$$

$$Where P = \text{phenanthrene}$$

$$1\text{-MP} = 1\text{-methylphenanthrene}$$

$$2\text{-MP} = 2\text{-methylphenanthrene}$$

$$3\text{-MP} = 3\text{-methylphenanthrene}$$

$$9\text{-MP} = 9\text{-methylphenanthrene}$$

$$1.5\text{-DMN} = 1.5\text{-dimethylnaphthalene}$$

$$2.6\text{-DMN} = 2.6\text{-dimethylnaphthalene}$$

$$2.7\text{-DMN} = 2.7\text{-dimethylnaphthalene}$$

Peak areas measured from m/z 156 (dimethylnaphthalene), m/z 178 (phenanthrene) and m/z 192 (methylphenanthrene) mass fragmentograms of diaromatic and triaromatic hydrocarbon fraction isolated by thin layer chromatography.

Recalibration of the methylphenanthrene index using data from a suite of Australian coals has given rise to another equation for calculated vitrinite reflectance (after Boreham et al, 1988):

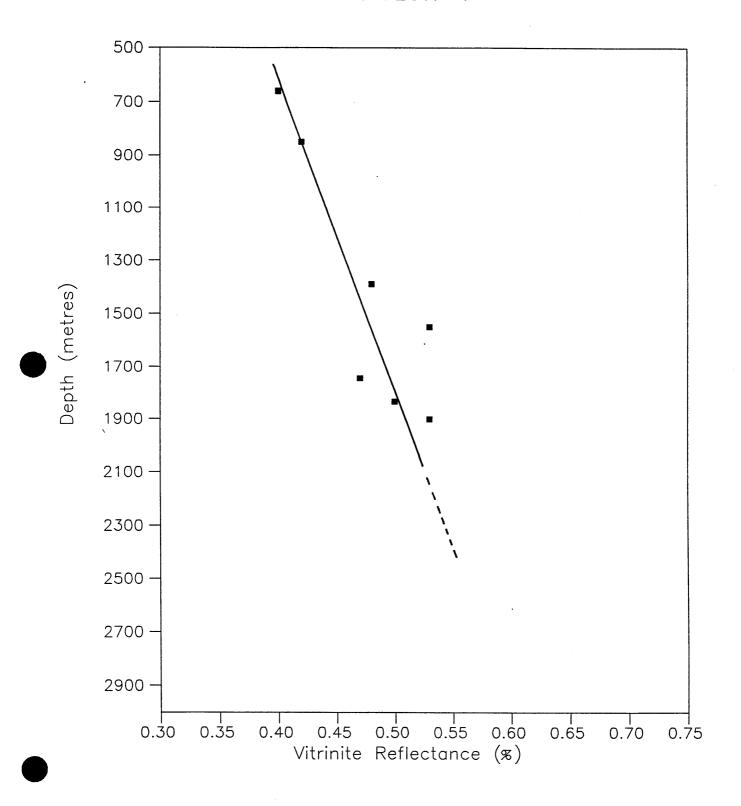
$$VR_{calc}$$
 (e) = 0.7 MPI + 0.22 (for VR < 1.7%)

The methylphenanthrene distribution ratio (MPDF) and calculated vitrinite reflectance VR<sub>calc</sub> (f) is derived from the following equation (after Kvalheim *et al*, 1987):

MPDF = 
$$\frac{(2-MP + 3-MP)}{(2-MP + 3-MP + 1-MP + 9-MP)}$$
  
VR<sub>calc</sub> (f) = -0.166 + 2.242 MPDF

VITRINITE REFLECTANCE VERSUS DEPTH MYLOR-1

FIGURE 1



### HYDROGEN INDEX vs T max

Client: Bridge Oil Limited Location: Mylor—1

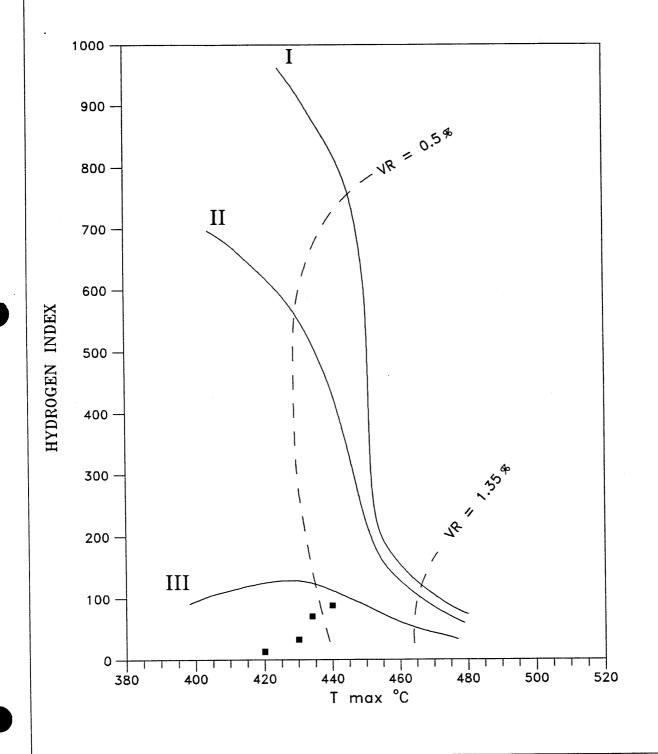
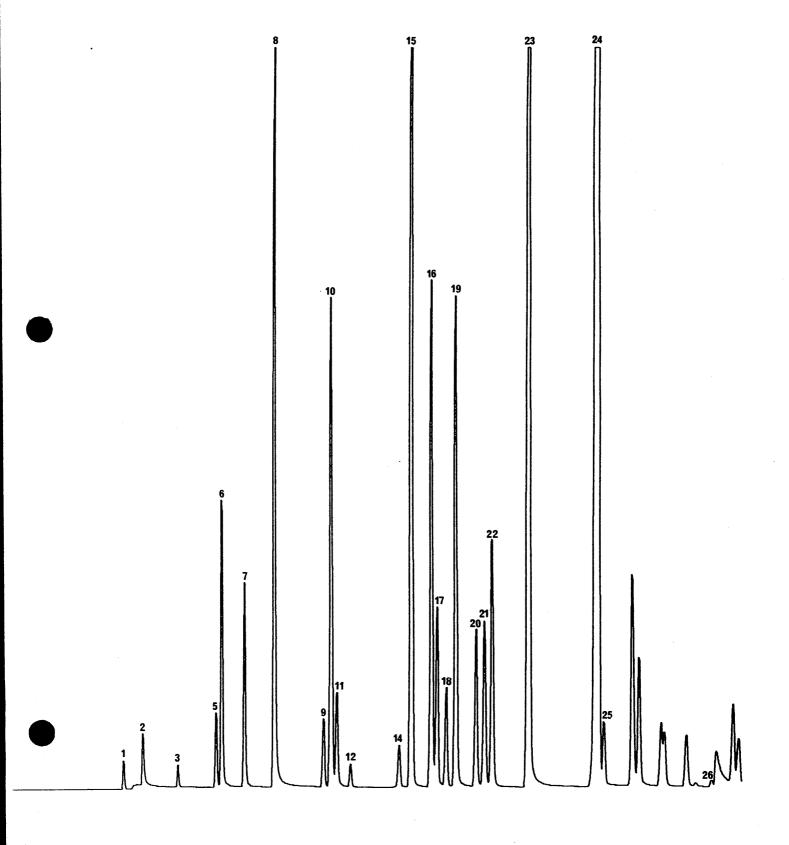


FIGURE 3

Mylor-1 DST-1 Condensate Gasoline Range Chromatogram



### **KEY TO GASOLINE-RANGE CHROMATOGRAM**

- 1. 2-Methylbutane (Isopentane)
- 2. n-Pentane
- 3. 2,2-Dimethylbutane
- 4. Cyclopentane
- 5. 2,3-Dimethylbutane
- 6. 2-Methylpentane
- 7. 3-Methylpentane
- 8. n-Hexane
- 9. 2,2-Dimethylpentane
- 10. Methylcyclopentane
- 11. 2,4-Dimethylpentane
- 12. 2,2,3-Trimethylbutane
- 13. Benzene
- 14. 3,3-Dimethylpentane
- 15. Cyclohexane
- 16. 2-Methylhexane
- 17. 2,3-Dimethylpentane
- 18. 1,1-Dimethylcyclopentane
- 19. 3-Methylhexane
- 20. cis-1,3-Dimethylcyclopentane
- 21. trans-1,3-Dimethylcyclopentane
- 22. 3-Ethylpentane and trans-1,2-Dimethylcyclopentane
- 23. *n*-Heptane
- 24. Methylcyclohexane
- 25. Ethylcyclopentane
- 26. Toluene

FIGURE 4

OIL MATURITY AND ALTERATION MYLOR-1

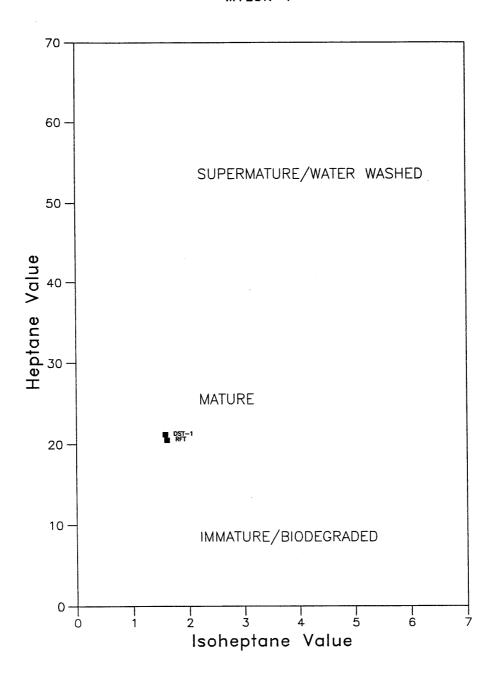
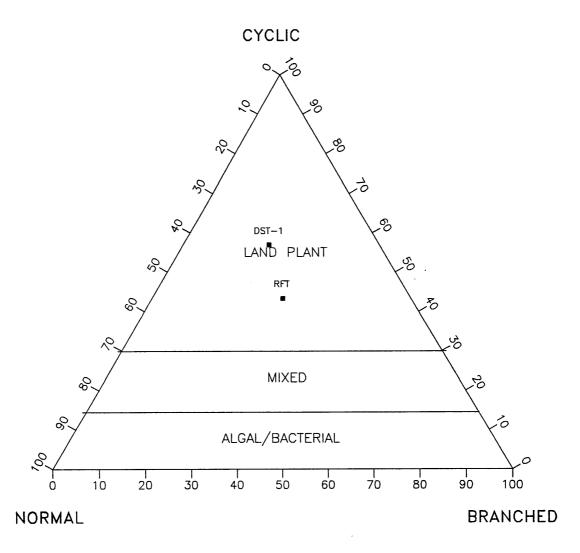


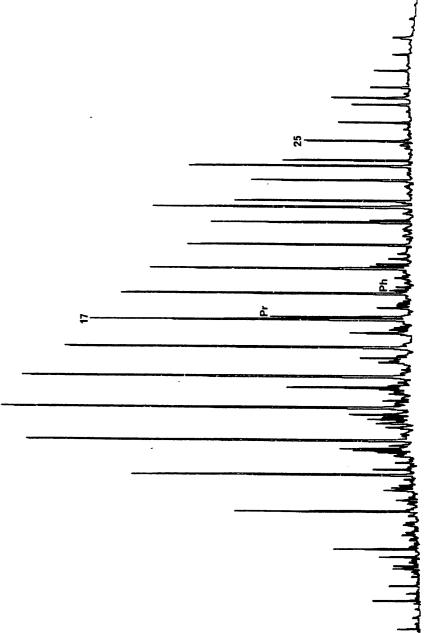
FIGURE 5

### OIL SOURCE AFFINITY BASED ON $C_5-C_7$ ALKANES MYLOR-1



# FIGURE 6

Mylor-1 Core Sample 1685.74-1685.86 metres GC of Extractable Organic Matter



Mylor-1 Core Sample 1702.9 metres GC of Extractable Organic Matter FIGURE 7 1

# FIGURE 8

Mylor-1
DST-1, Condensate
GC of saturates fraction

The state of the s

FIGURE 9

MYLOR-1 GENETIC AFFINITY AND MATURITY

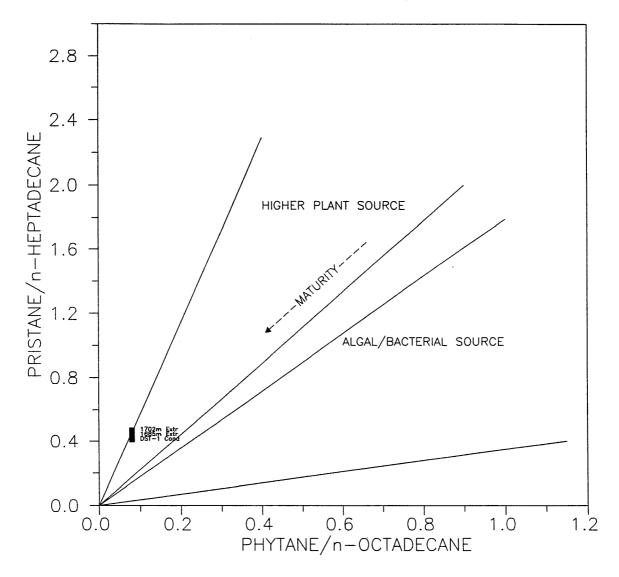
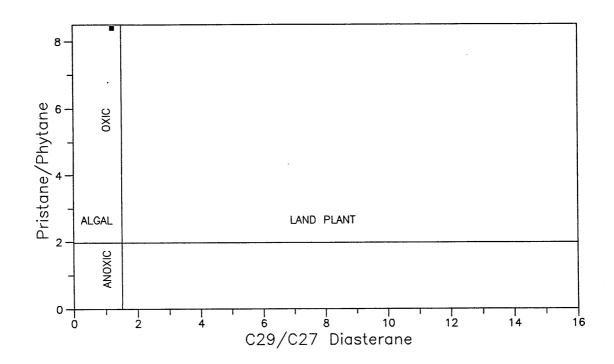


FIGURE 10

### MYLOR-1,DST-1 OIL SOURCE AFFINITY



C<sub>29</sub> STERANE MATURITY - MIGRATION PLOT MYLOR-1, DST-1

FIGURE 11

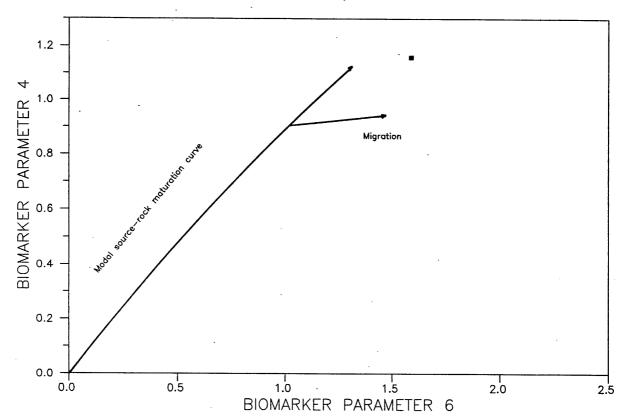


FIGURE 12

STERANE DISTRIBUTIONS MYLOR-1, DST-1

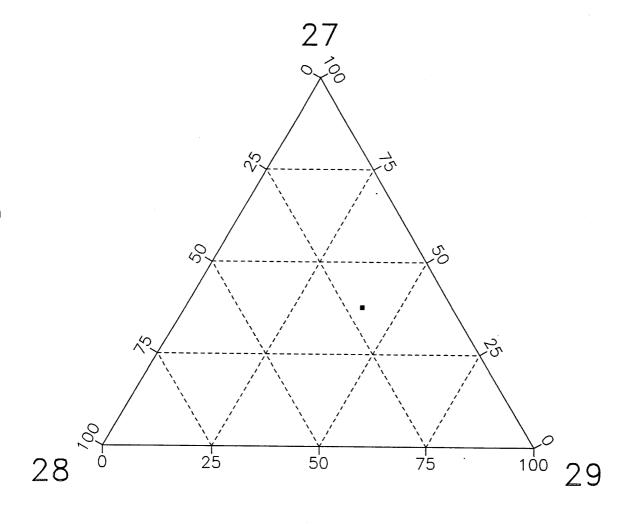
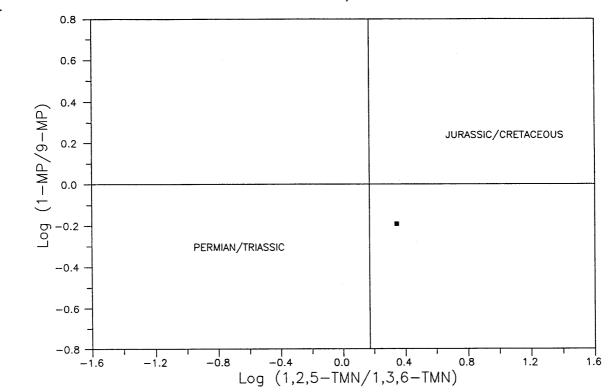


FIGURE 13

### AROMATIC BIOMARKERS IN OIL MYLOR-1, DST-1



## APPENDIX 1

#### **ANALYTICAL METHODS**

#### 1. Sample Preparation

Samples (as received) were ground in a Siebtechnik mill for 20-30 seconds.

#### 2. Total Organic Carbon (TOC)

Total organic carbon was determined by digestion of a known weight (approximately 0.2 g) of powdered rock in HCl to remove carbonates, followed by combustion in oxygen in the induction furnace of a Leco WR-12 Carbon Determinator and measurement of the resultant CO<sub>2</sub> by infra-red detection.

#### 3. Rock-Eval Pyrolysis

A 100 mg portion of powdered rock was analysed by the Rock-Eval pyrolysis technique (Girdel IFP-Fina Mark 2 instrument; operating mode, Cycle 1).

#### 4. Organic Petrology

Representative portions of each sample (crushed to -14+35 BSS mesh) were obtained with a sample splitter and then mounted in cold setting Glasscraft resin using a 2.5 cm diameter mould. Each block was ground flat using diamond impregnated laps and carborundum paper. The surface was then polished with aluminium oxide and finally magnesium oxide.

Reflectance measurements were made with a Leitz MPV1.1 microphotometer fitted to a Leitz Ortholux microscope and calibrated against synthetic standards. All measurements were taken using oil immersion (n = 1.518) and incident monochromatic light (wavelength 546 nm) at a temperature of 23±1°C. Fluorescence observations were made on the same microscope utilising a 3 mm BG3 excitation filter, a TK400 dichroic mirror and a K510 suppression filter.

#### 5. Isolation of Residual Oil

Core chips and cuttings samples were extracted with dichloromethane in Soxhlet apparatus for 8 hours. Removal of solvent by careful rotary evaporation gave the oil (nominal  $C_{12+}$  fraction).

#### 6. Liquid Chromatography

Asphaltenes were not precipitated from the condensate prior to liquid chromatography. The samples were separated into hydrocarbons (saturates and aromatics) and polar compounds (resins) by liquid chromatography on activated alumina (sample: adsorbent ratio = 1.100). Hydrocarbons were eluted with petroleum ether/dichloromethane (75:25) and resins with methanol/dichloromethane (65:35).

The saturated and aromatic hydrocarbons were then separated by liquid chromatography on activated silica gel (sample/adsorbent ratio = 1:100). The saturated hydrocarbons were eluted with petroleum ether and the aromatic hydrocarbons with petroleum ether/dichloromethane (91:9).

#### 7. Gas Chromatography

Whole oils and saturated hydrocarbons (alkanes) were examined by gas chromatography using the following instrumental parameters:

Gas Chromatograph:

Perkin Elmer 8500 operated in the split

injection mode

Column:

25 m x 0.3 mm fused silica, SGE

QC3/BP1

**Detector Temperature:** 

300°C

Column Temperature:

40°C for 1 minute, then 8° per minute

to 300°C and held isothermal at 300°C

until all peaks eluted

Quantification:

Relative concentrations of individual hydrocarbons were obtained by measurement of peak areas with a Perkin-Elmer LCI 100 integrator. The areas of peaks responding to aromatic hydrocarbons were multiplied by

appropriate response factors

#### 8. Thin Layer Chromatography (TLC)

Aromatic hydrocarbons were isolated from the extracted oil by preparative TLC using Merck GF<sub>254</sub> silica plates and distilled AR grade n-pentane as eluent. Naphthalene and anthracene were employed as reference standards for the diaromatic and triaromatic hydrocarbons, respectively. These two bands, visualised under UV light, were scraped from the plate and the aromatic hydrocarbons redissolved in dichloromethane.

GC-MS analysis of the aromatic hydrocarbons was undertaken in the selected ion detection (SID) mode. The instrument and its operating parameters were as follows:

System: Perkin-Elmer 8420 GC coupled with a

Finigan Ion Trap mass selective detector

and data system

Column: 25 mm x 0.2 mm i.d. HP BP5 cross-

linked methylsilicone phase fused silica, interfaced to source of mass

spectrometer

Injector: Split injection (8:1)

Carrier Gas: He at 60 Kpa head pressure

Column Temperature: 50-260 °C @ 4 °/minute

Mass Spectrometer

Conditions: Selected ion monitoring

The following mass fragmentograms were recorded:

m/z	Compound Type
155 + 156	dimethylnaphthalenes
169 + 170	trimethylnaphthalenes
178	phenanthrene
191 + 192	methylphenanthrene

The area of the phenanthrene peak was multiplied by a response factor of 0.667 when calculating the methylphenanthrene index (MPI).

Naphthenes (branched/cyclic alkanes) were isolated from the oil by molecular sieve separation of the saturates fraction.

GC-MS analysis of the naphthenes was undertaken in the multiple ion detection (MID) mode. Instrumental conditions are given below.

System:	HP	5890	Series	II	Plus	GC	coupled	to
---------	----	------	--------	----	------	----	---------	----

HP 5972 MSD

Column: 25 mm x 0.25 mm i.d. HPS MS cross-

linked methylsilicone phase fused silica, interfaced directly to source of mass

spectrometer

Injector: Splitless  $2\mu$ L

Carrier Gas: Helium at a linear velocity of 26

cm/minute

Column Temperature: 50°C for 2 minutes then 50-290°C @

7°/minute

Mass Spectrometer

Conditions: 70 eV EI; 9-ion selected ion monitoring,

70 millisec dwell time for each ion

The following mass fragmentograms were recorded:

m/z	Compound Type
83	alkylcyclohexanes
123	drimanes, diterpanes
177	demethylated triterpanes
183	acyclic alkanes (incl isoprenoids,
	botryococcanes)
191	triterpanes (incl hopanes, moretanes)
205	methyltriterpanes
217	steranes
218	steranes
231	4-methylsteranes
259	diasteranes

#### 10. Gasoline-Range Hydrocarbons

The condensate samples were analysed on a Perkin-Elmer 8500 Gas Chromatograph equipped with a 50 m, 0.2 mm i.d. HP PONA column. Concentrations of  $C_3$  -  $C_8$  hydrocarbons were calculated from the peak areas for each compound.

## **APPENDIX 2**

## VITRINITE REFLECTANCE VALUES AND HISTOGRAMS

Well:

Mylor-1

Depth:

650-660 m.

## Sorted List

0.39

0.40

Number of values 2 Mean of values 0.40 Standard Deviation 0.01

HISTOGRAM OF VALUES
Reflectance values multiplied by 100

39-41 \*\*

Well:

Mylor-1

Depth:

840-850 m.

## Sorted List

0.40

0.41

0.44

Number of values 3 Mean of values 0.42 Standard Deviation 0.02

# HISTOGRAM OF VALUES Reflectance values multiplied by 100

40-42 \*\* 43-45 \*

Well: Depth:

Mylor-1

th: 1386-1389 m.

# 0.43 0.49 0.45 0.49 0.45 0.50 0.46 0.50 0.47 0.50

0.510.51

0.51

0.52

0.47

0.48

0.48

0.48

Standard Deviation

Sorted List

Number of values 20 Mean of values 0.48

HISTOGRAM OF VALUES
Reflectance values multiplied by 100

0.02

43-45 \*\*\*

46-48 \*\*\*\*\*\*

49-51 \*\*\*\*\*\*\*

52-54 \*

Well: Depth:

Mylor-1

1550 m.

#### Sorted List

0.51

0.51

0.52

0.52

0.54

0.57

Number of values6Mean of values0.53Standard Deviation0.02

# HISTOGRAM OF VALUES Reflectance values multiplied by 100

51-53 \*\*\*\*

54-56 \*

57-59 \*

Well: Depth: Mylor-1 1744 m.

## Sorted List

0.40	0.48
0.41	0.50
0.42	0.50
0.43	0.50
0.44	0.51
0.46	0.52
0.46	
0.47	
0.47	
0.48	

Number of values	16
Mean of values	0.47
Standard Deviation	0.04

# HISTOGRAM OF VALUES Reflectance values multiplied by 100

```
40-42 ***

43-45 **

46-48 *****

49-51 ****

52-54 *
```

Well: Depth:

Mylor-1 1835 m.

#### Sorted List

0.43	0.50
0.43	0.51
0.47	0.51
0.47	0.52
0.47	0.53
0.48	0.55
0.49	0.55
0.50	0.57
0.50	
0.50	

Number of values 18
Mean of values 0.50
Standard Deviation 0.04

# HISTOGRAM OF VALUES

Reflectance values multiplied by 100

```
43-45 **
46-48 ****
49-51 ******
52-54 **
55-57 ***
```

Well:

Mylor-1

Depth: 1901 m.

#### Sorted List

0.52

0.52

0.52

0.54

0.54

0.55

Number of values 6
Mean of values 0.53
Standard Deviation 0.01

# HISTOGRAM OF VALUES Reflectance values multiplied by 100

52-54 \*\*\* 55-57 \*\*\*

## **APPENDIX 3**

## **PHOTOMICROGRAPHS**

#### PE906752

This is an enclosure indicator page.

The enclosure PE906752 is enclosed within the container PE900935 at this location in this document.

The enclosure PE906752 has the following characteristics:

ITEM\_BARCODE = PE906752
CONTAINER\_BARCODE = PE900935

NAME = Photomicrograph, Appendix 5, Figure 1-2

BASIN =

OTWAY

PERMIT = PEP108

TYPE = WELL

SUBTYPE = PHOTOMICROGRAPH

DESCRIPTION = Photomicrograph, Appendix 5, Figure

1-2, Mylor-1

REMARKS =

 $DATE\_CREATED = 18/10/94$ 

 $DATE\_RECEIVED = 23/03/95$ 

 $W_NO = W1102$ 

WELL\_NAME = MYLOR-1 CONTRACTOR = AMDEL

CLIENT\_OP\_CO = BRIDGE OIL LIMITED

(Inserted by DNRE - Vic Govt Mines Dept)

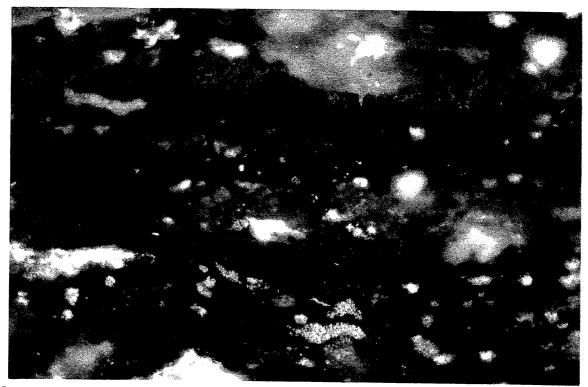


Plate 1: 1744 metres depth

Reflected Light

Vitrinite (grey) occurs in stringers in this sample. Inertinite (light grey) is present as inertodetrinite.

Field dimensions: 0.26 x 0.53 mm.



Plate 2: 1744 metres depth

Fluorescence Mode

Resinite (intense orange) cutinite and liptodetrinite (moderate orange) are more easily distinguished in fluorescence mode and are associated with the vitrinite in this field of view.

PE906752

#### PE906753

This is an enclosure indicator page.

The enclosure PE906753 is enclosed within the container PE900935 at this location in this document.

The enclosure PE906753 has the following characteristics:

ITEM\_BARCODE = PE906753
CONTAINER\_BARCODE = PE900935

NAME = Photomicrograph, Appendix 5, Figure 3-4

BASIN =

OTWAY

PERMIT = PEP108

TYPE = WELL

 $\mathtt{SUBTYPE} = \mathtt{PHOTOMICROGRAPH}$ 

DESCRIPTION = Photomicrograph, Appendix 5, Figure

3-4, Mylor-1

REMARKS =

DATE\_CREATED = 18/10/94

DATE\_RECEIVED = 23/03/95

 $W_NO = W1102$ 

WELL\_NAME = MYLOR-1

CONTRACTOR = AMDEL

CLIENT\_OP\_CO = BRIDGE OIL LIMITED

(Inserted by DNRE - Vic Govt Mines Dept)

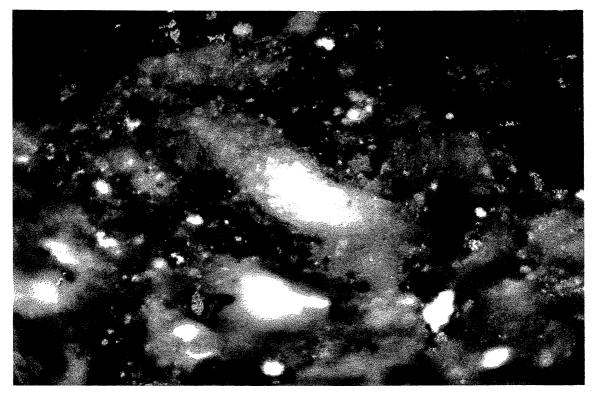


Plate 3: 1744 metres depth

Reflected Light

Organic matter in this SWC consists largely of inertinite (grey). Small detrital vitrinite is also present in this sample.

Field dimensions: 0.26 x 0.53 mm.

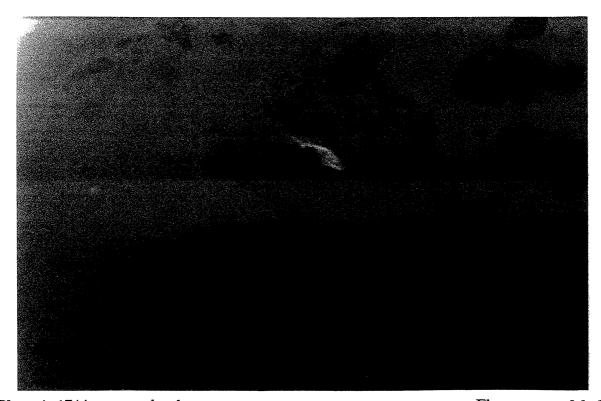


Plate 4: 1744 metres depth

Fluorescence Mode

Phytoplankton and litodetrinite are identified in fluorescence mode.



#### PE906754

This is an enclosure indicator page.

The enclosure PE906754 is enclosed within the container PE900935 at this location in this document.

The enclosure PE906754 has the following characteristics:

ITEM\_BARCODE = PE906754
CONTAINER\_BARCODE = PE900935

NAME = Photomicrograph, Appendix 5, Figure 5-6
BASIN =

OTWAY

PERMIT = PEP108 TYPE = WELL

SUBTYPE = PHOTOMICROGRAPH

DESCRIPTION = Photomicrograph, Appendix 5, Figure

5-6, Mylor-1

REMARKS =

DATE\_CREATED = 18/10/94

DATE\_RECEIVED = 23/03/95

 $W_NO = W1102$ 

WELL\_NAME = MYLOR-1 CONTRACTOR = AMDEL

CLIENT\_OP\_CO = BRIDGE OIL LIMITED

(Inserted by DNRE - Vic Govt Mines Dept)

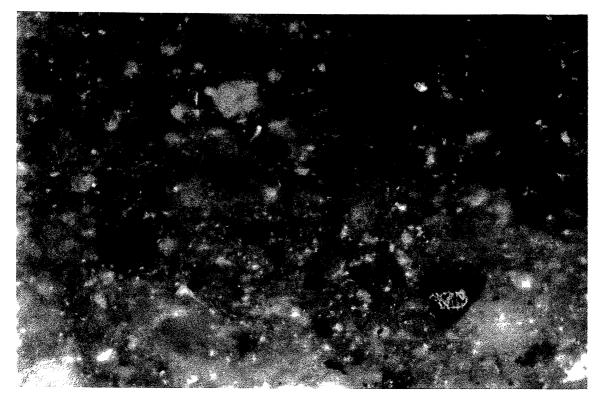


Plate 5: 1835 metres depth

Reflected Light

Organic matter in this field of view consists largely of detrital inertinite (inertodetrinite).

Field dimensions: 0.26 x 0.53 mm.



Plate 6: 1835 metres depth

Fluorescence Mode

The low abundance of liptinite is evident in this plate with only trace liptodetrinite being present.

DEPT. NAT. RES & ENV

PE906754

## **APPENDIX 4**

GC-MS OF BRANCHED/CYCLIC ALKANES

C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:

AMDEL-597

Method File:

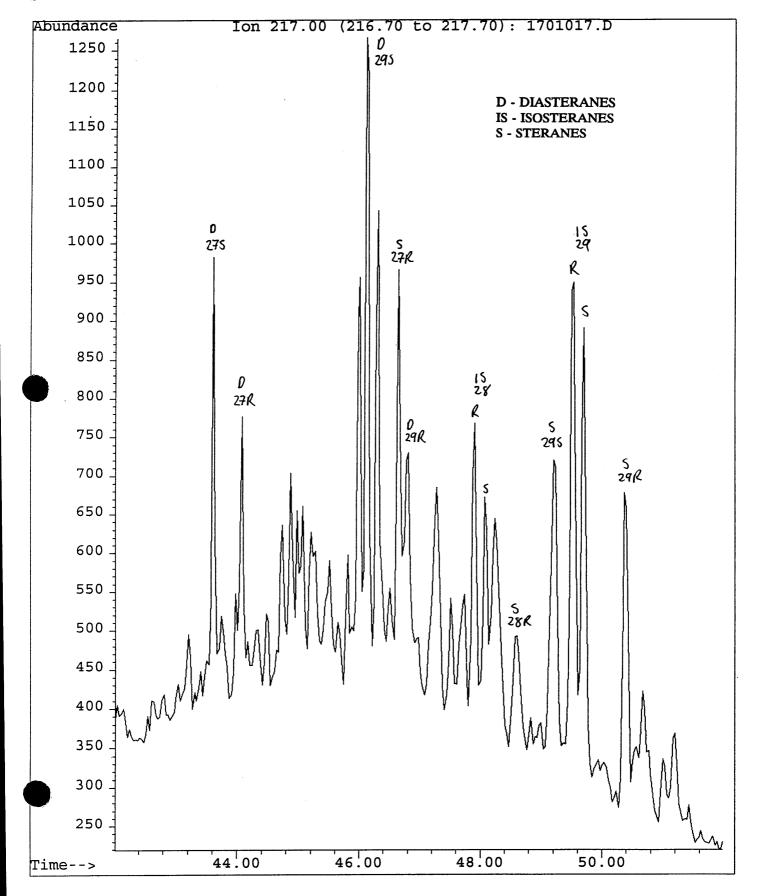
NAPHTH

Sample Name:

MYLOR-1, DST-1 CONDENSATE

isc Info:

ial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:
Method File:

AMDEL-597

Method File:

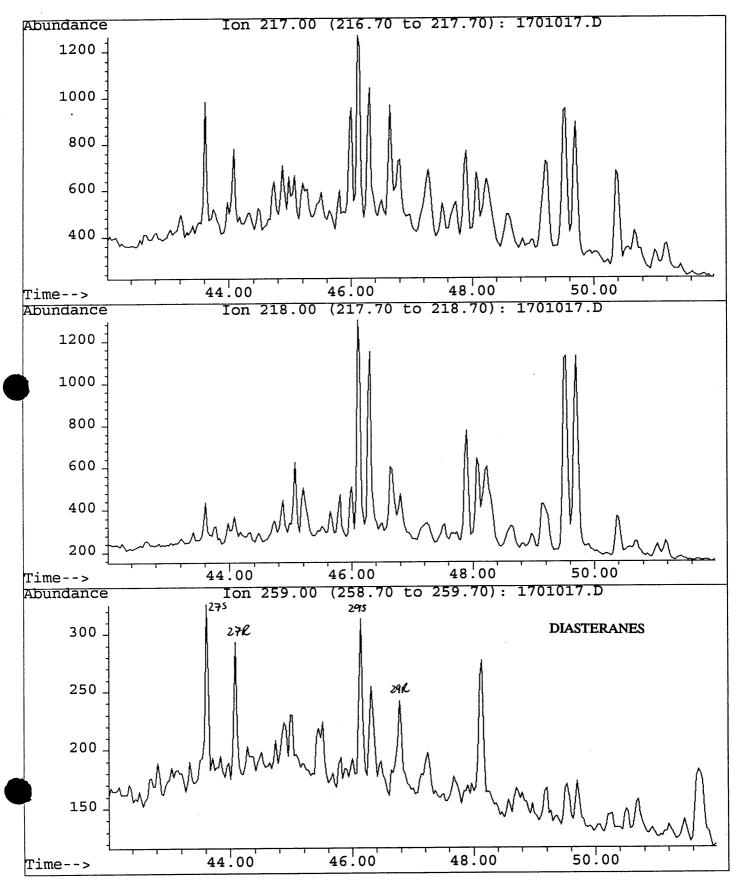
NAPHTH

Sample Name:

MYLOR-1, DST-1 CONDENSATE

Misc Info:

Vial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:
Method File:

AMDEL-597

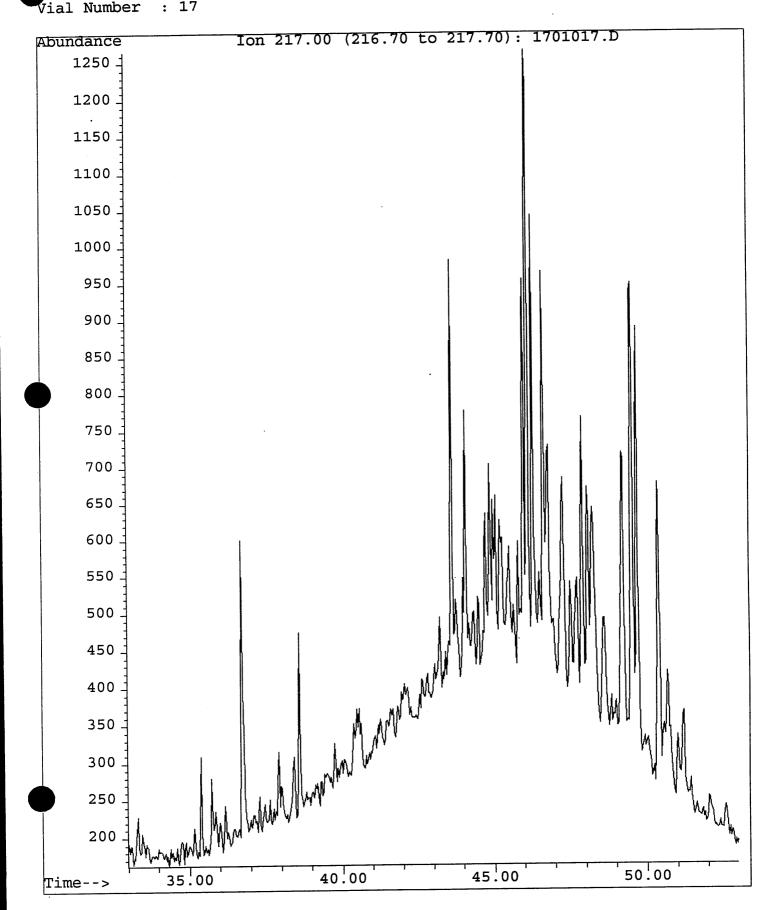
Sample Name:

NAPHTH

isc Info:

ist inito.

MYLOR-1, DST-1 CONDENSATE



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:
Method File:

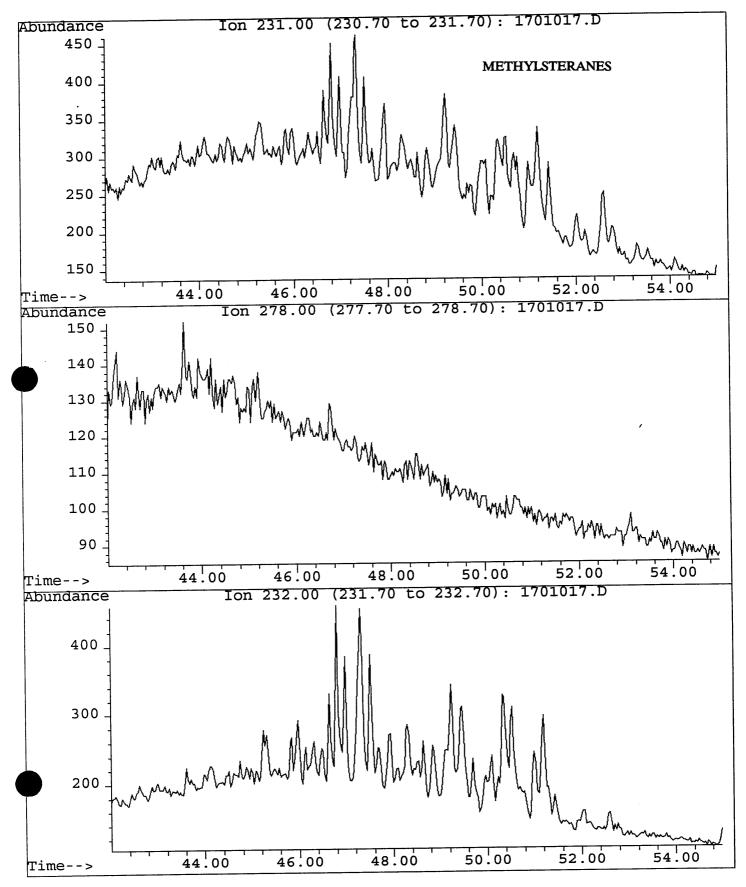
AMDEL-597

Method File: NAPA Sample Name:

NAPHTH
MYLOR-1, DST-1 CONDENSATE

isc Info:

Vial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument: Method File: AMDEL-597

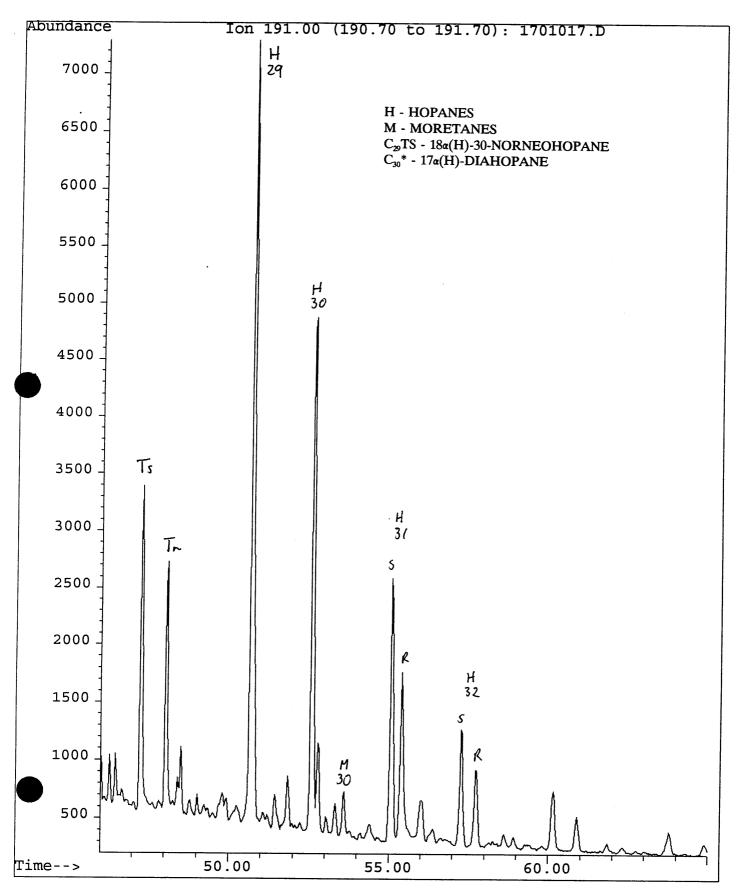
Sample Name:

NAPHTH

MYLOR-1, DST-1 CONDENSATE

isc Info:

ial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired: 6 Aug 94 10:42 am

Instrument:

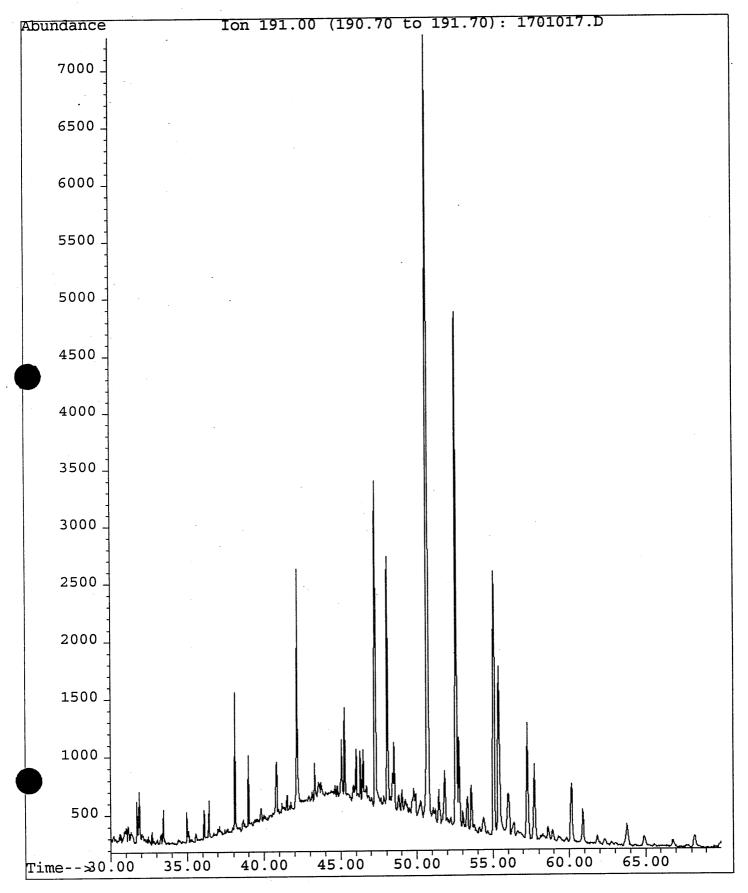
AMDEL-597

Method File: Sample Name: NAPHTH

MYLOR-1, DST-1 CONDENSATE

sc Info:

Vial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired: 6 Aug 94

10:42 am

Instrument:

AMDEL-597

Method File:

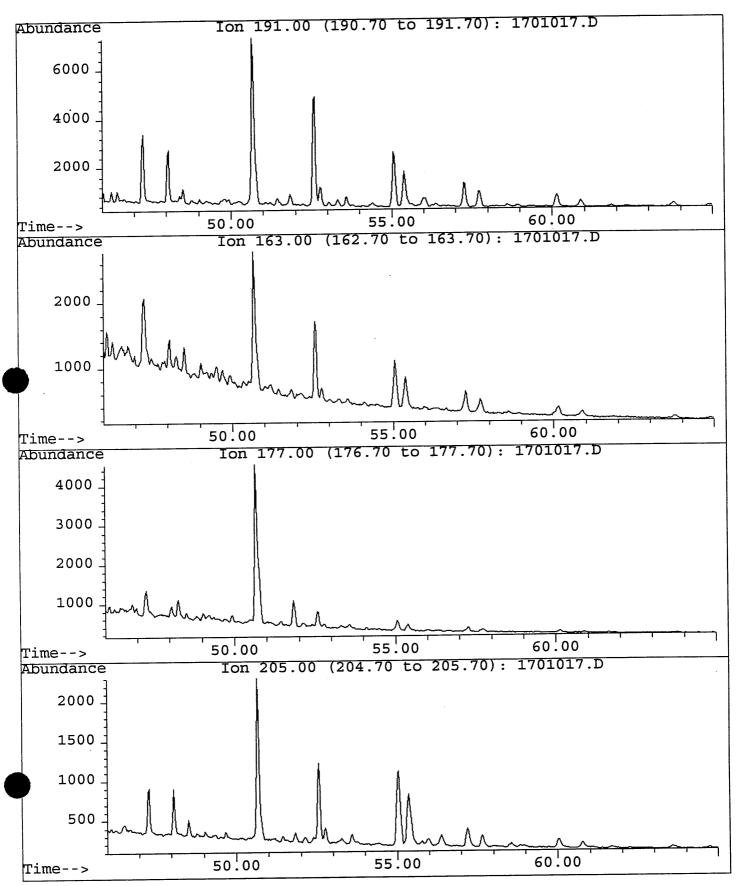
NAPHTH

Sample Name:

MYLOR-1, DST-1 CONDENSATE

isc Info:

Vial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

10:42 am 6 Aug 94

Instrument: Method File: AMDEL-597

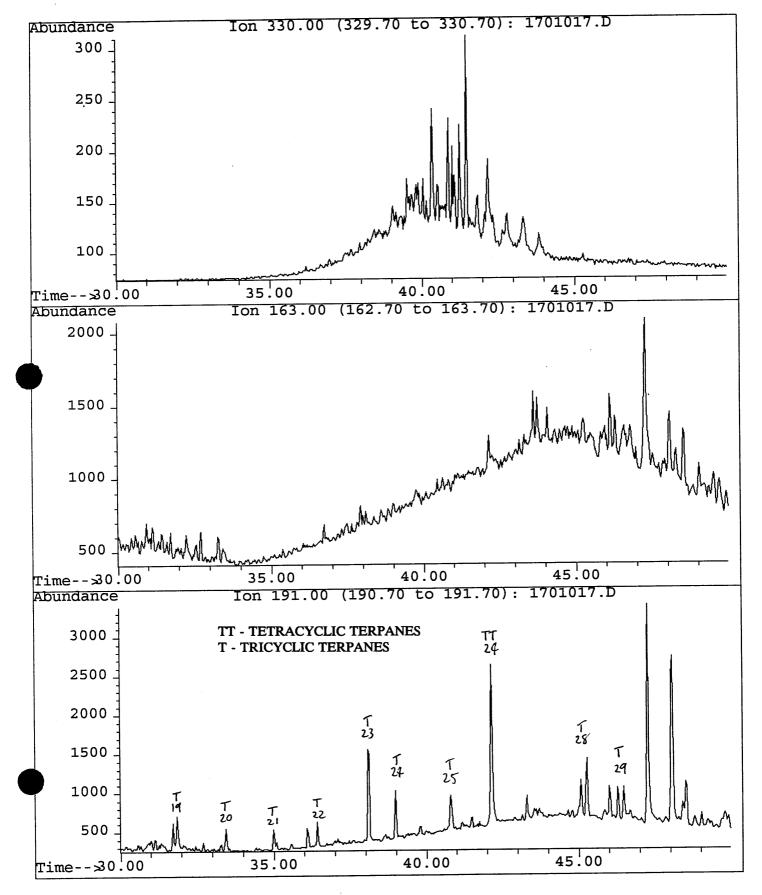
Sample Name:

NAPHTH

MYLOR-1, DST-1 CONDENSATE

isc Info:

Vial Number : 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:

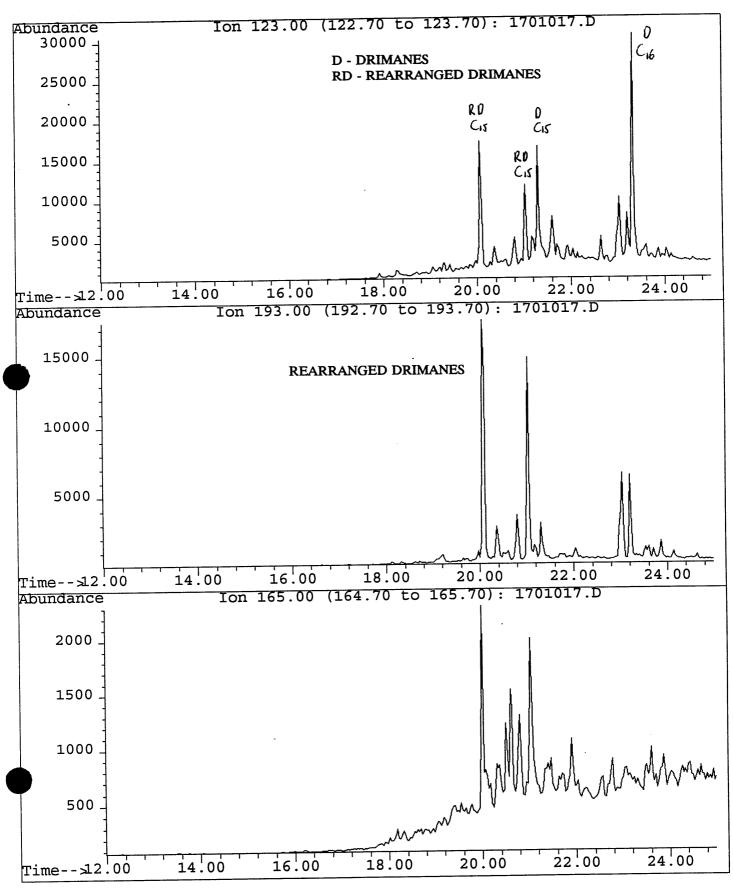
AMDEL-597 NAPHTH

Method File: Sample Name:

MYLOR-1, DST-1 CONDENSATE

isc Info:

Vial Number : 17



File : C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

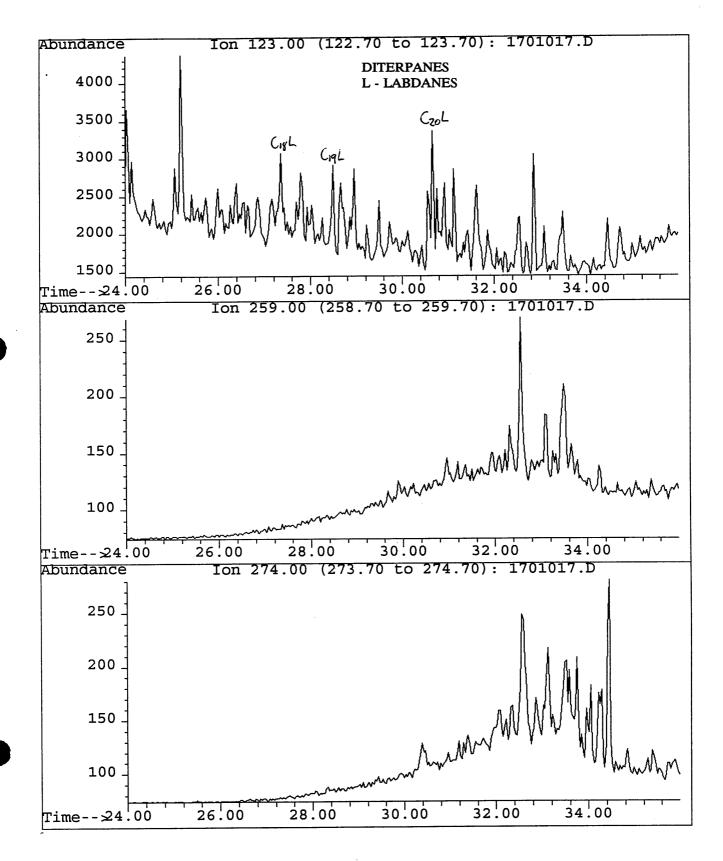
Operator : Scott Wythe

Acquired : 6 Aug 94 10:42 am using AcqMethod NAPHTH

Instrument: AMDEL-597

Sample Name: MYLOR-1, DST-1 CONDENSATE

Misc Info : Vial Number: 17



File : C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

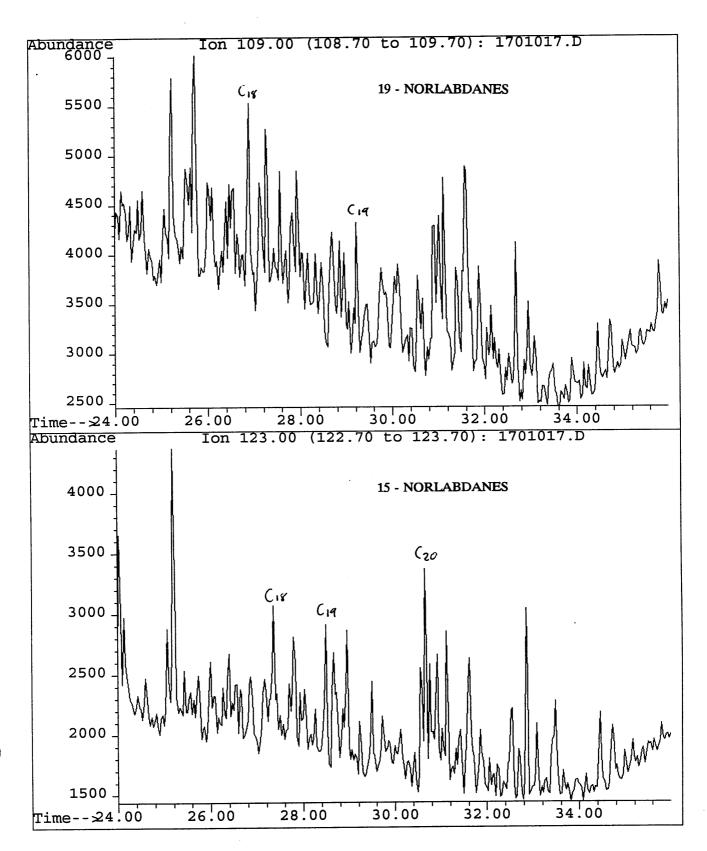
Operator : Scott Wythe

Acquired : 6 Aug 94 10:42 am using AcqMethod NAPHTH

Instrument: AMDEL-597

Sample Name: MYLOR-1, DST-1 CONDENSATE

Misc Info : Vial Number: 17



C:\HPCHEM\1\DATA\PETSERV\050894\1701017.D

Operator:

Scott Wythe

Date Acquired:

6 Aug 94 10:42 am

Instrument:

AMDEL-597

Method File:

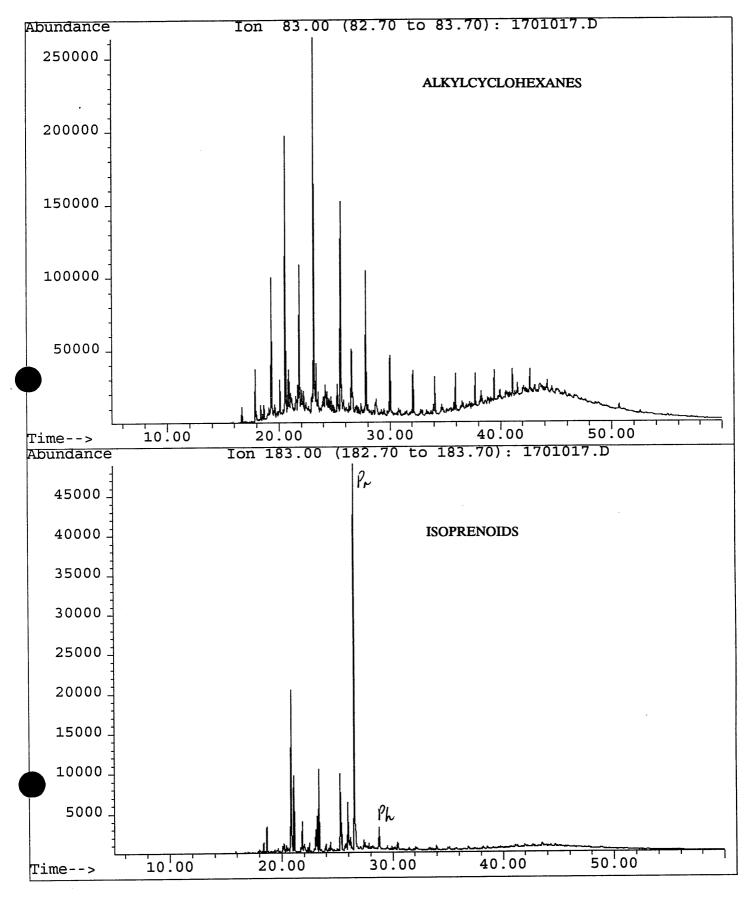
NAPHTH

Sample Name:

MYLOR-1, DST-1 CONDENSATE

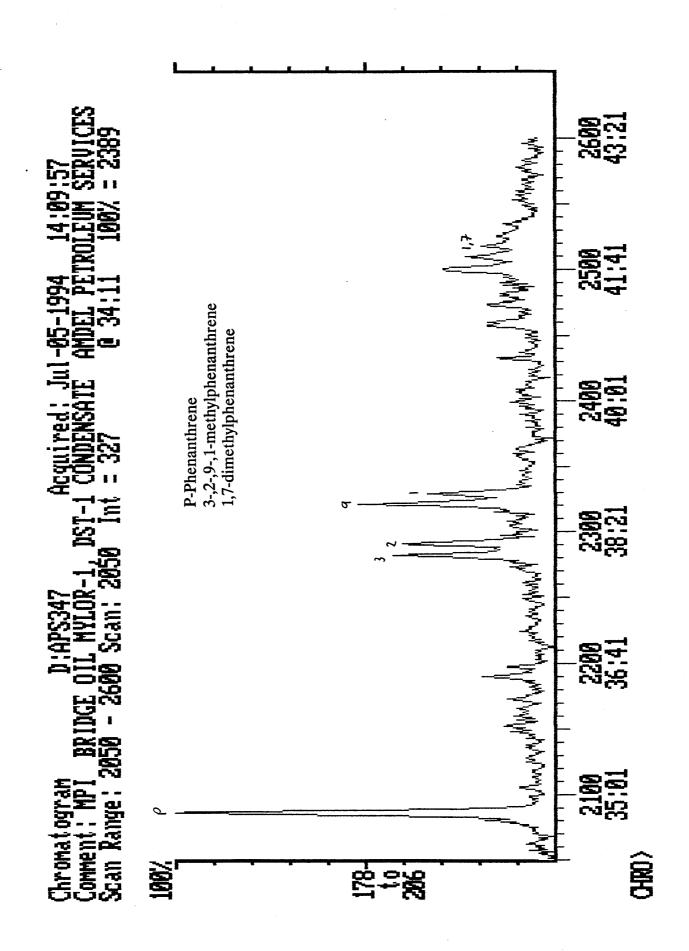
isc Info:

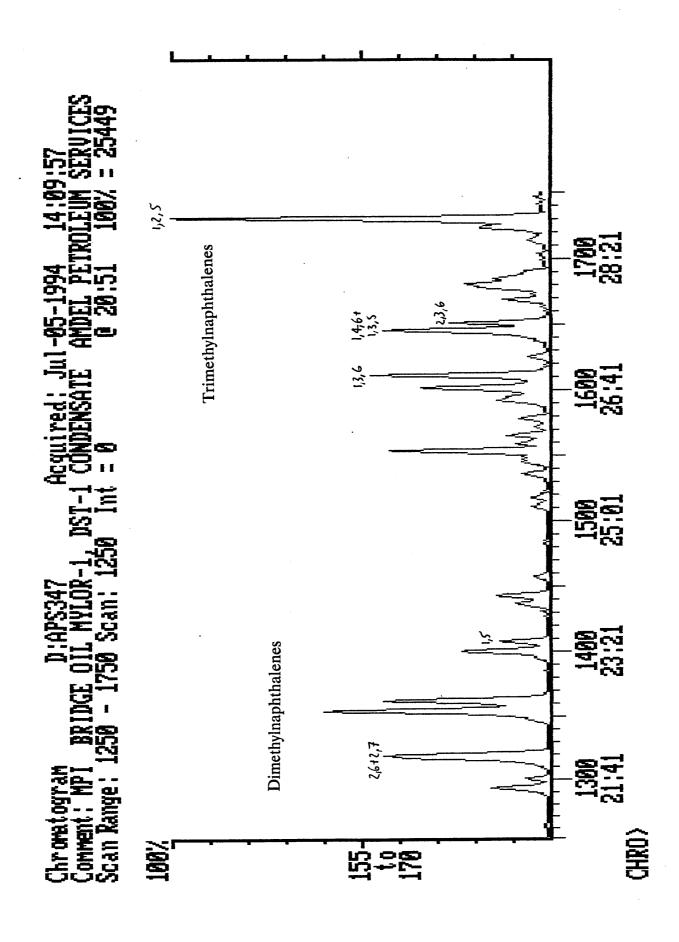
Vial Number : 17



## **APPENDIX 5**

## GC-MS OF AROMATIC HYDROCARBONS





# ENCLOSURES

# **ENCLOSURES**

1. Mud Log

PE 600746

#### PE600746

This is an enclosure indicator page. The enclosure PE600746 is enclosed within the container PE900935 at this location in this document.

The enclosure PE600746 has the following characteristics:

ITEM\_BARCODE = PE600746
CONTAINER\_BARCODE = PE900935

NAME = Formation Evaluation Log/Mud Log

BASIN = OTWAY PERMIT = PEP 108

TYPE = WELL

SUBTYPE = WELL\_LOG

DESCRIPTION = XL Base Formation Evaluation/ Mud Log

(enclosure 1 from WCR vol.2) for

Mylor-1

REMARKS =

DATE\_CREATED = 24/06/94

DATE\_RECEIVED =

W\_NO = W1102 WELL\_NAME = Mylor-1

CONTRACTOR = Baker Hughes Inteq CLIENT\_OP\_CO = Bridge Oil Ltd

(Inserted by DNRE - Vic Govt Mines Dept)

# **ENCLOSURES**

2. Core Photographs

PE900936 PE 906755

#### PE900936

This is an enclosure indicator page.

The enclosure PE900936 is enclosed within the container PE900935 at this location in this document.

The enclosure PE900936 has the following characteristics:

ITEM\_BARCODE = PE900936
CONTAINER\_BARCODE = PE900935

NAME = Core Photographs

BASIN = OTWAY
PERMIT = PEP 108
TYPE = WELL

 $SUBTYPE = CORE_PHOTOS$ 

DESCRIPTION = Core Photographs - (enclosure 2 from

WCR vol.2) for Mylor-1

REMARKS = The photographs have been stuck together to form a core log.

DATE\_CREATED =

DATE\_RECEIVED =

 $W_NO = W1102$ 

WELL\_NAME = Mylor-1

CONTRACTOR = Bridge Oil Limited CLIENT\_OP\_CO = Bridge Oil Ltd

(Inserted by DNRE - Vic Govt Mines Dept)

#### PE906755

This is an enclosure indicator page. The enclosure PE906755 is enclosed within the container PE900935 at this location in this document.

The enclosure PE906755 has the following characteristics:

ITEM\_BARCODE = PE906755
CONTAINER\_BARCODE = PE900935

NAME = UV Core Photographs

BASIN = OTWAY
PERMIT = PEP108
TYPE = WELL

SUBTYPE = CORE\_PHOTOS

DESCRIPTION = UV Core Photographs for Mylor-1

REMARKS = Plastic Strip of Photos

DATE\_CREATED =

DATE\_RECEIVED = 23/03/95

 $W_NO = W1102$ 

WELL\_NAME = MYLOR-1

CONTRACTOR =

CLIENT\_OP\_CO = BRIDGE OIL LIMITED

(Inserted by DNRE - Vic Govt Mines Dept)

# **ENCLOSURES**

3. Composite Log

PE 600747

#### PE600747

This is an enclosure indicator page.

The enclosure PE600747 is enclosed within the container PE900935 at this location in this document.

The enclosure PE600747 has the following characteristics:

ITEM\_BARCODE = PE600747

CONTAINER\_BARCODE = PE900935

NAME = Composite Log

BASIN = OTWAY

PERMIT = PEP 108

TYPE = WELL

SUBTYPE = COMPOSITE\_LOG

DESCRIPTION = Composite Log (enclosure 3 from WCR

vol.2) for Mylor-1

REMARKS =

DATE CREATED = 28/06/94

DATE\_RECEIVED =

 $W_NO = W1102$ 

WELL\_NAME = Mylor-1

CONTRACTOR = Parker & Parsley Australasia Ltd

CLIENT\_OP\_CO = Bridge Oil Ltd

(Inserted by DNRE - Vic Govt Mines Dept)

# **ENCLOSURES**

4. Mylor #1 Discovery Montage

PE900937

#### PE900937

This is an enclosure indicator page.

The enclosure PE900937 is enclosed within the container PE900935 at this location in this document.

The enclosure PE900937 has the following characteristics:

ITEM\_BARCODE = PE900937
CONTAINER\_BARCODE = PE900935

NAME = Seismic Line - Discovery Montage

BASIN = OTWAY
PERMIT = PEP 108
TYPE = WELL
SUBTYPE = MONTAGE

DESCRIPTION = Seismic Line 7005, Trace 1795 Waarre 3D

- Discovery Montage (enclosure 4 from

WCR vol.2) for Mylor-1

REMARKS = Very Large Colour Seismic Chart

DATE\_CREATED =

DATE\_RECEIVED =

W\_NO = W1102 WELL\_NAME = Mylor-1

CONTRACTOR =

CLIENT\_OP\_CO = Bridge Oil Ltd

(Inserted by DNRE - Vic Govt Mines Dept)